

Fabrication and Imaging of 2D Nanomembranes and Graphene using

Outline:

Part I : Fabrication of 2D carbon nanostructures

Nanomembranes from SAMs

- Graphene and Graphenoids
- Nanoribbons and Nanosieves

Chemical Lithography

- Polymer Carpets
- Protein Biochips
- Janus Membranes

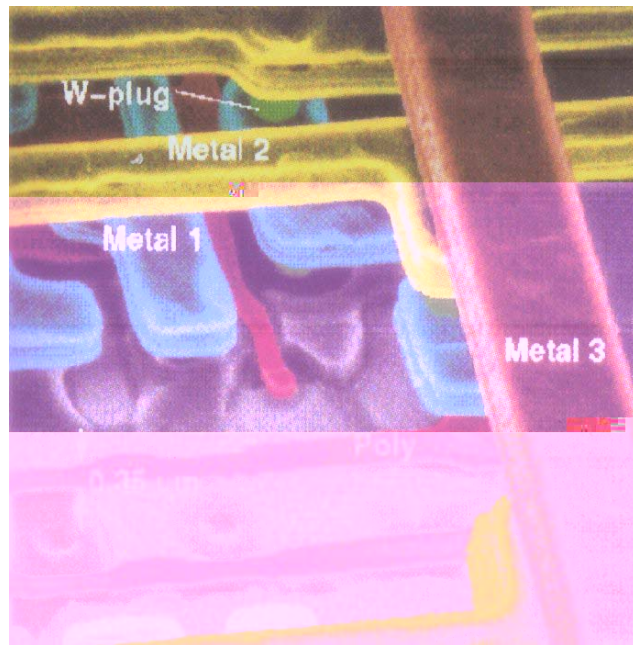
Part II : Helium Ion Microscopy

Basics

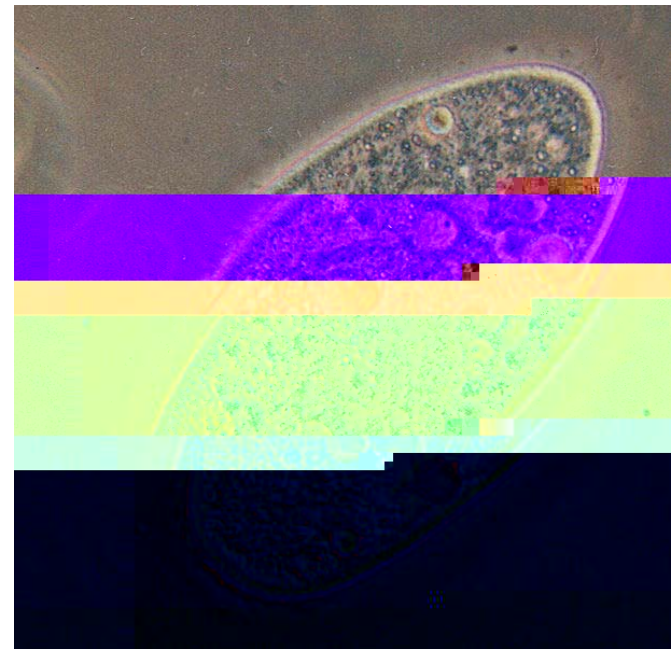
Nanomembranes and BioImaging

Concepts of Nanostructure Fabrication

**Integrated circuit
Lithography
(physics, engineering)**

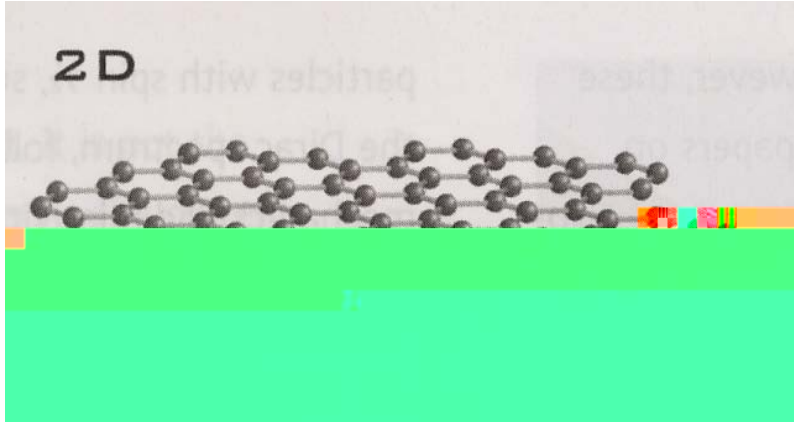


**Eukaryotic cell
Self-assembly
(chemistry, biology)**



Objective: Building (bio)functional molecular nanostructures with lithography and self-assembly

2-Dimensional Carbon Nanostructures



Graphene:

solid state, hard

Fabrication procedures :

- *Exfoliation of graphite/HOPG*
- *Epitaxy of SiC/TiC*
- *Oxidation/reduction of graphite*
- *CVD of hydrocarbons*

Hard to functionalize

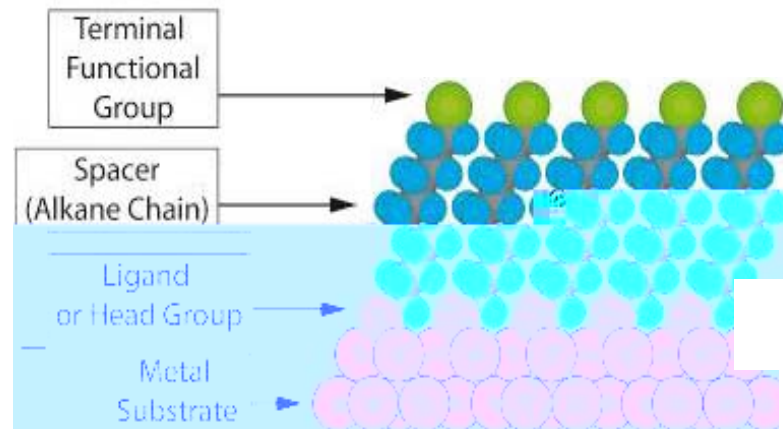
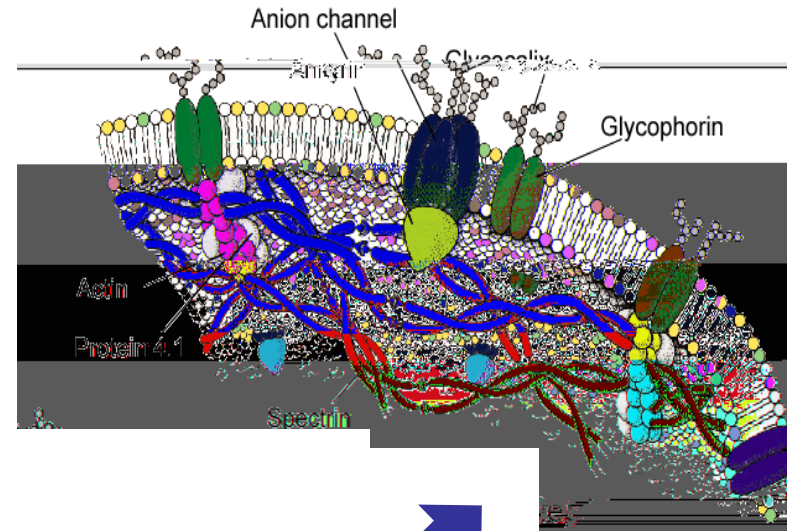
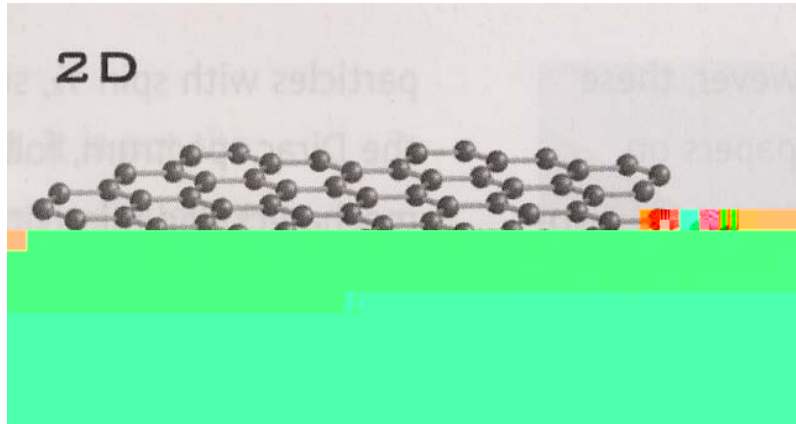
Cell membranes:

molecular, soft, directional

Fabrication procedures :

- *Self-Assembly of Tc T(4MC / Texb6T.*

2-Dimensional Carbon Nanostructures



Self-Assembled Monolayer (SAM):

molecular, soft, directional

Fabrication:

Surface chemistry, Intermolecular interactions, lateral ordering, 2D-crystallization

A molecular path to two-dimensional carbon nanostructures

Molecules

Solid substrates

self-assembly

Self-Assembled Monolayers (SAMs)

cross-linking by electron-beam

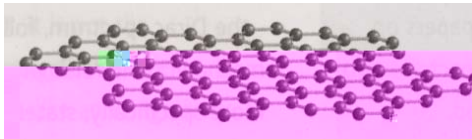
Carbon Nanomembranes

pyrolysis

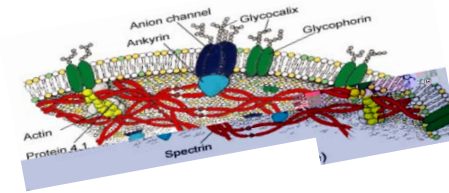
chemical, biological
functionalization

Graphene and Graphenoids

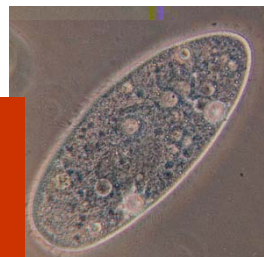
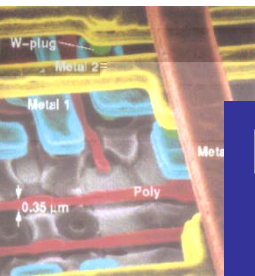
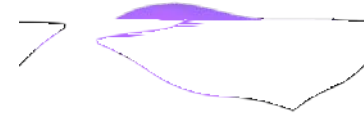
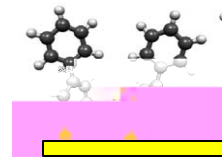
Functional Membranes



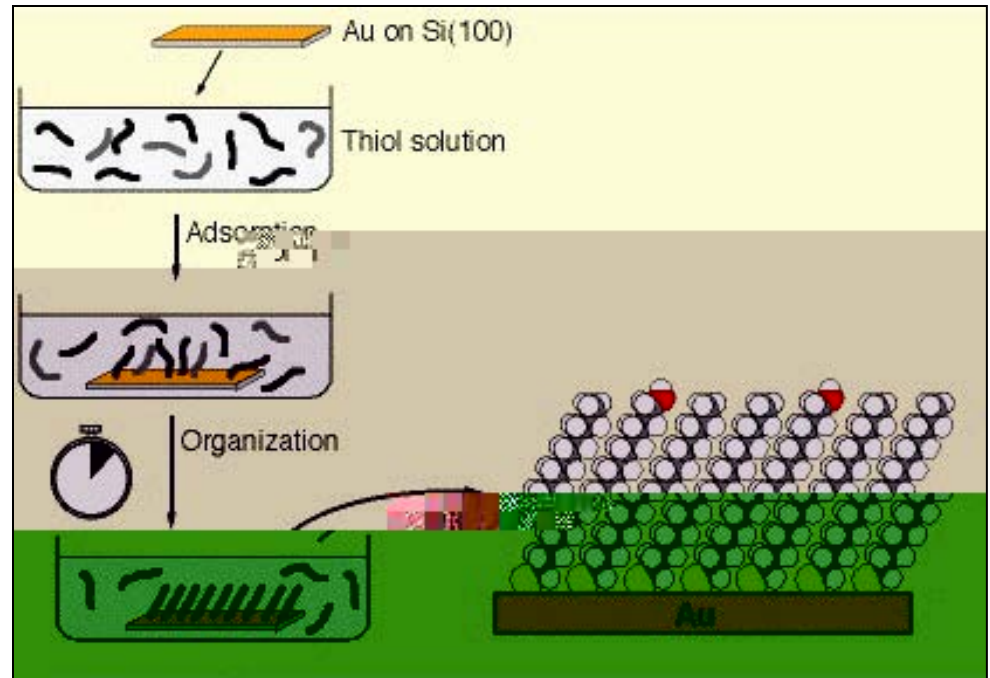
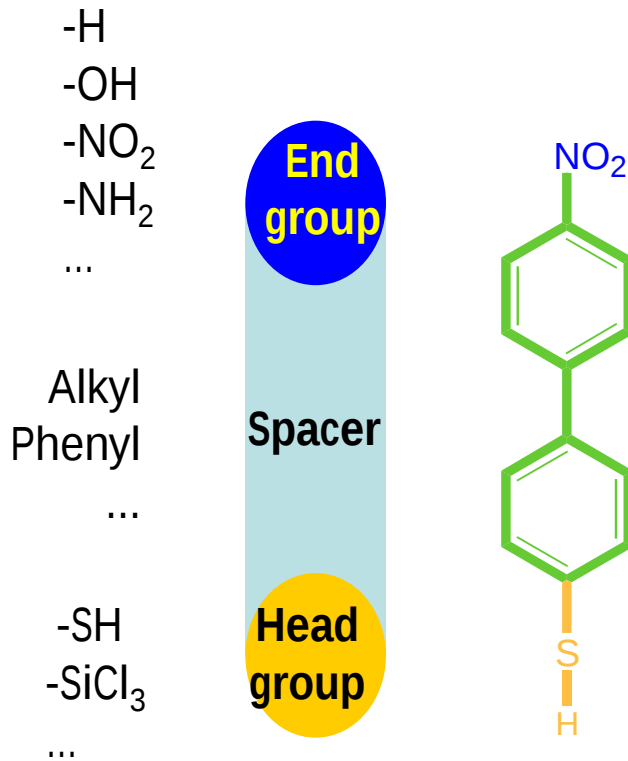
Electronics, NEMS,
Sensors, ...



Biomimetic, Medical,
Biosensors, ...



Self-Assembled Monolayers (SAMs)



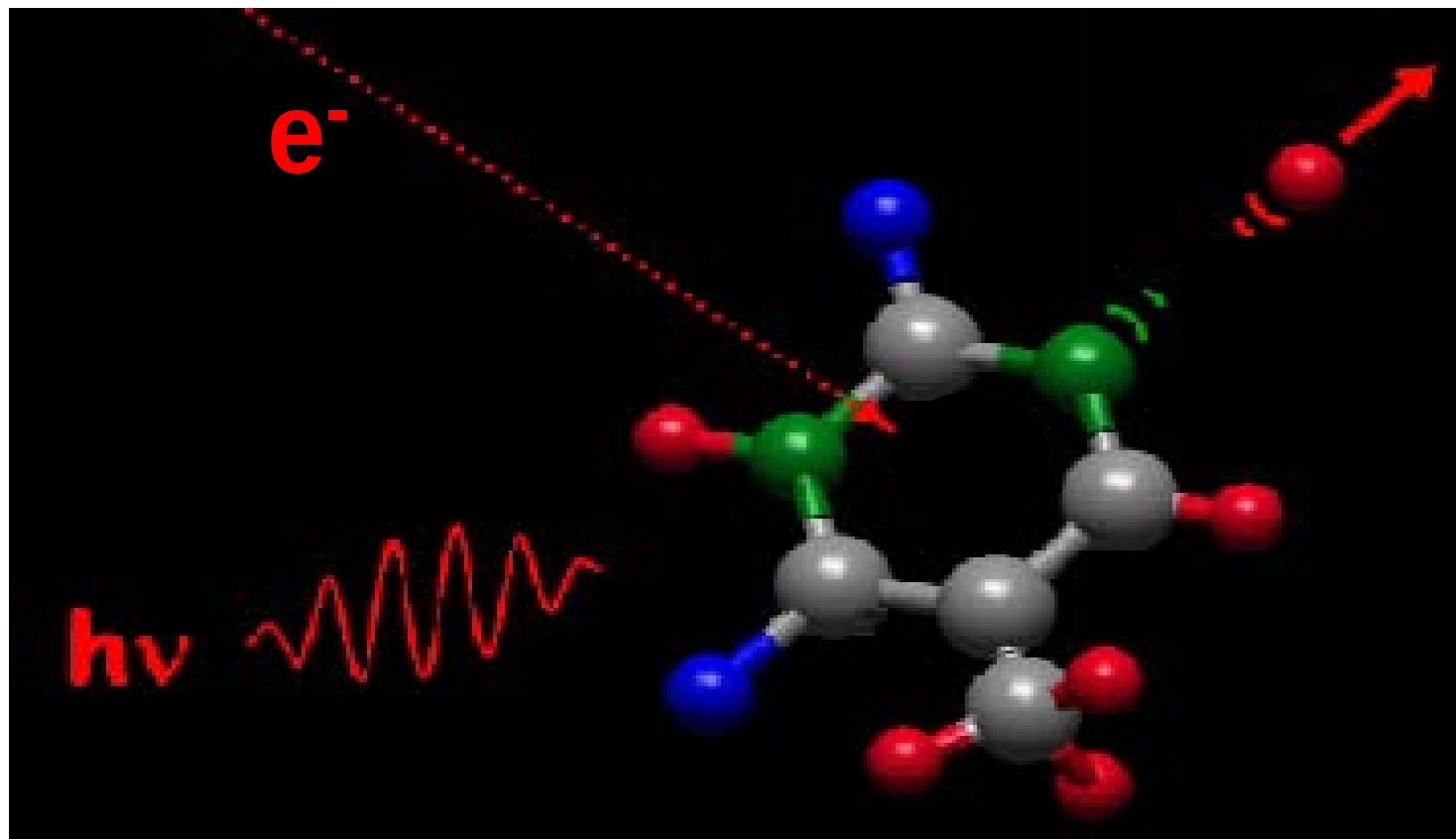
soft film on hard substrate

Fabrication procedures and conditions:

- *Liquid state, solutions*
- *ambient temperature and atmospheric pressure*
- *crystallization, equilibrium*

Easy to functionalize by choice of molecules and substrate

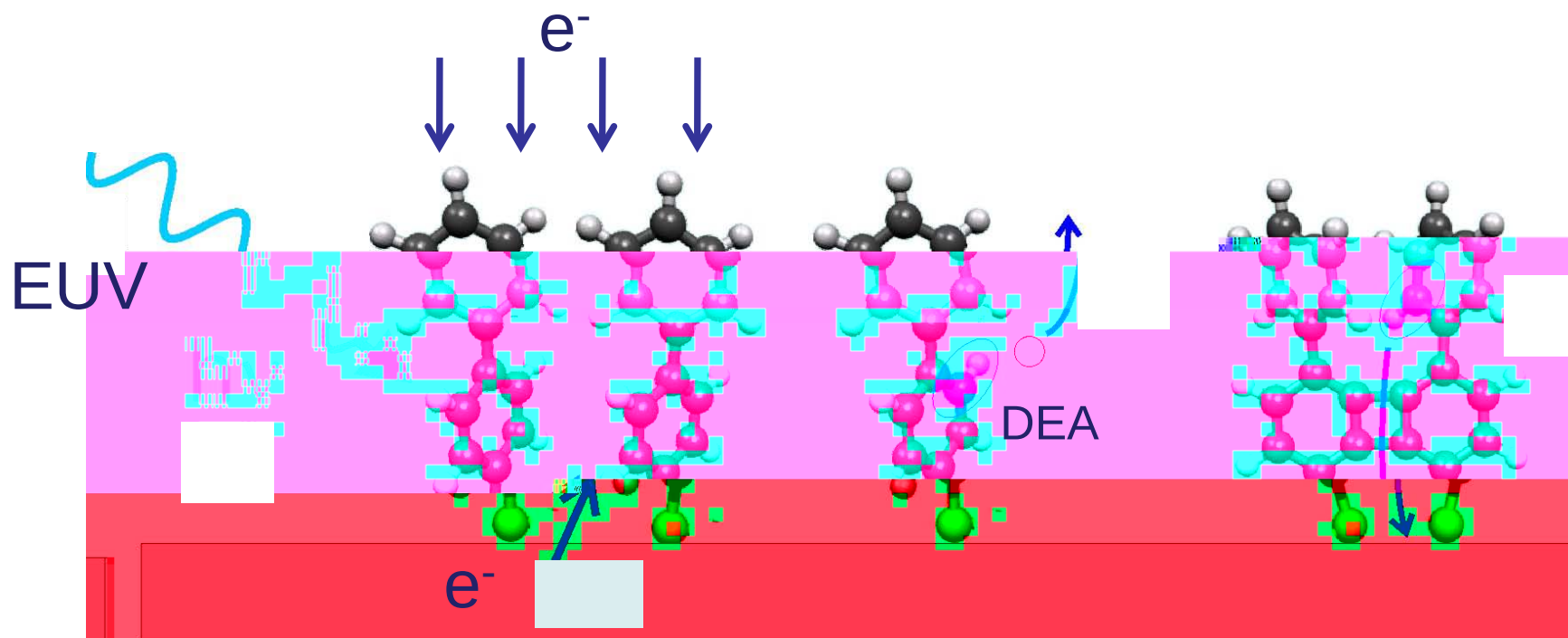
Electron and Photon induced Chemical Control



Electron-molecule interaction:

- ... Dissociative Electron Attachment (DEA) via Transient Negative Ion (TNI)
- ... requires low electron energies... typically below 10 eV

Electron and Photon induced Chemical Control in SAMs



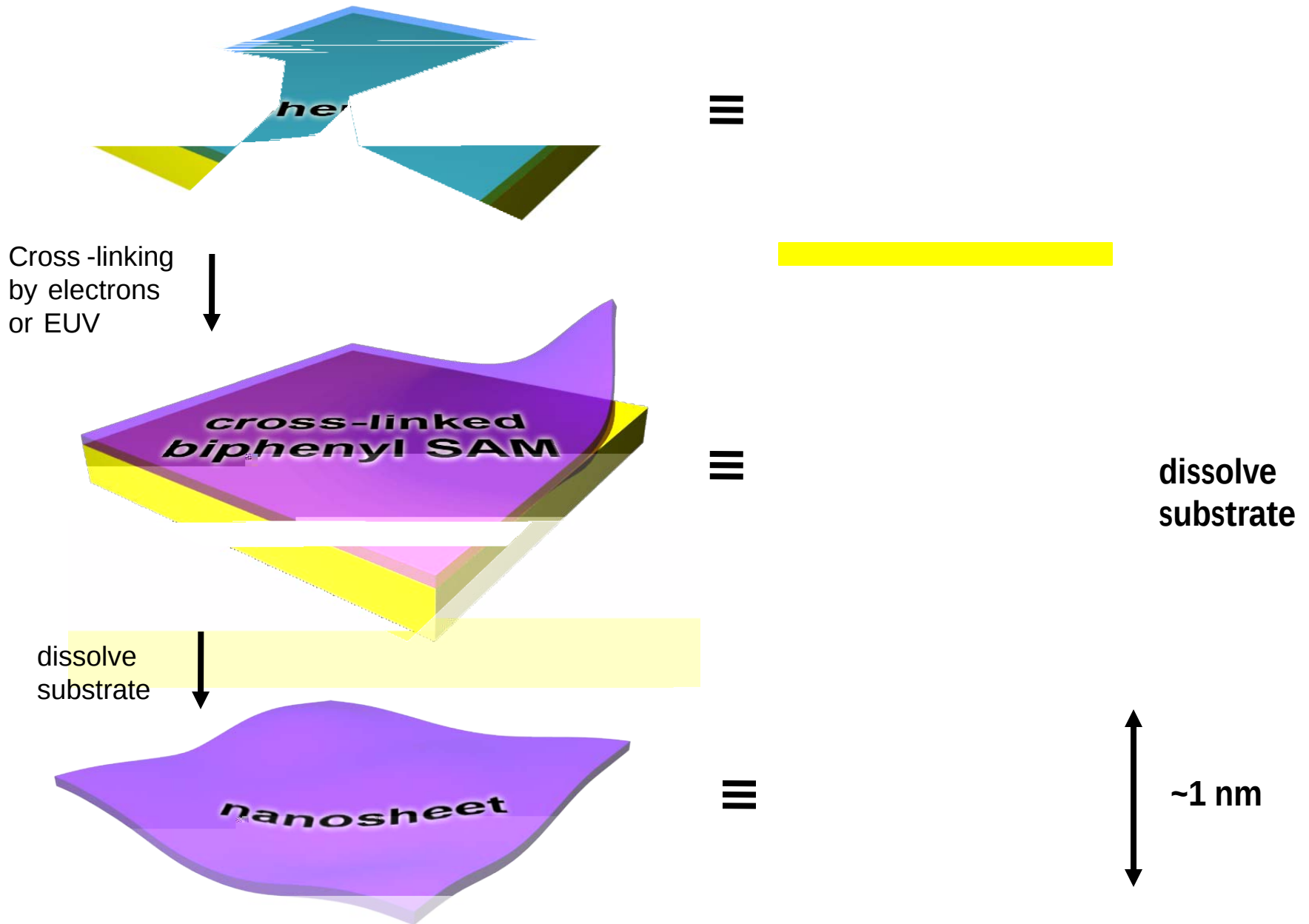
W. Geyer et al. *Appl. Phys. Lett* 75, 2401 (1999)

W. Eck et al. *Adv. Mater.* 12, 805 (2000)

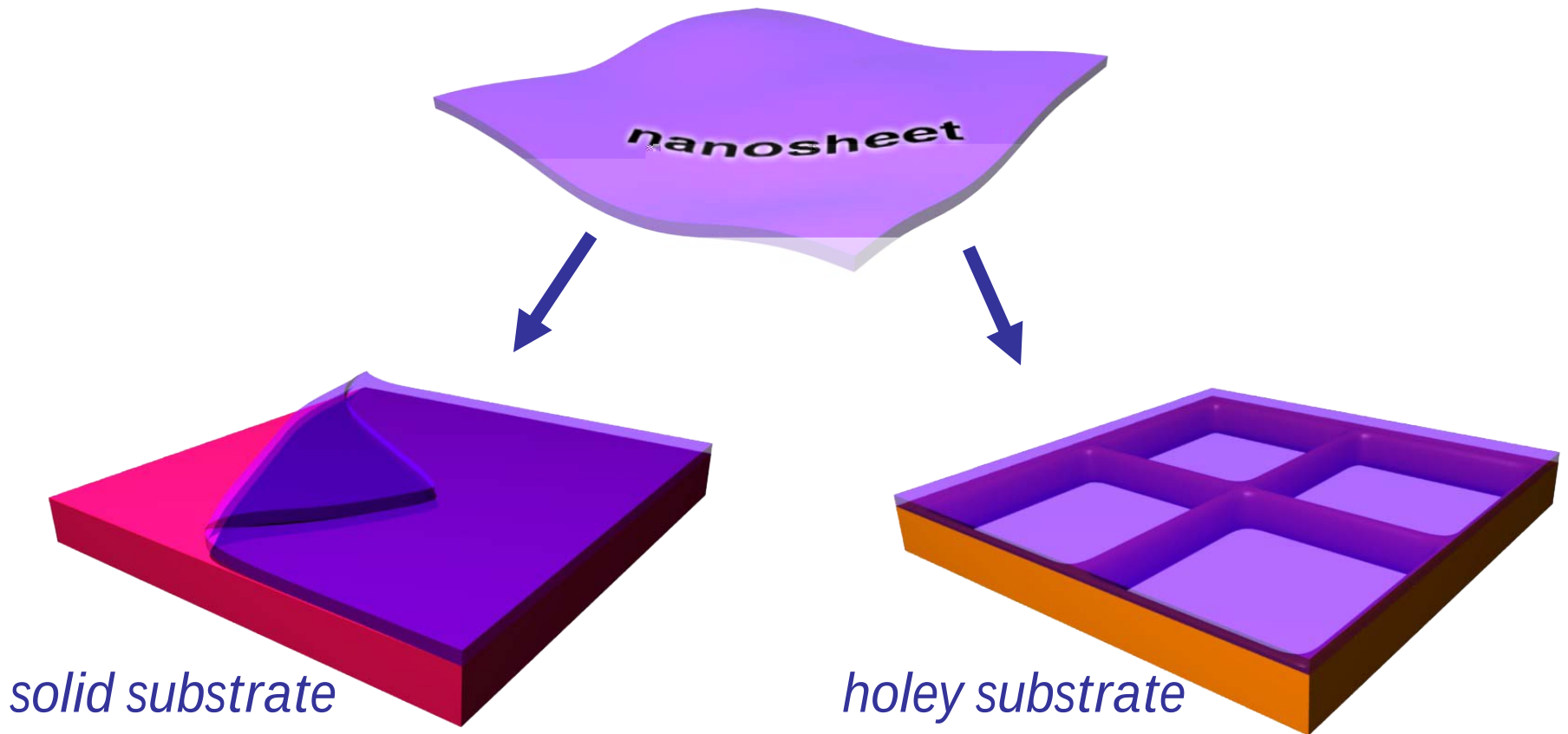
A. Turchanin et al. *Small* 3, 2114 (2007)

A. Turchanin et al. *Langmuir* 25, 7372 (2009)

Preparation of Nanomembrane



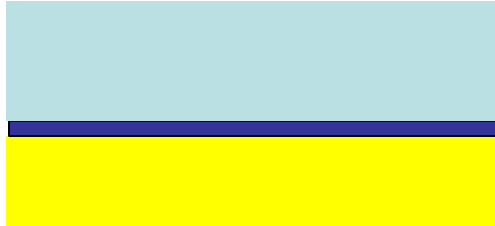
Preparation of Nanomembrane



Transfer of carbon nanomembrane, process



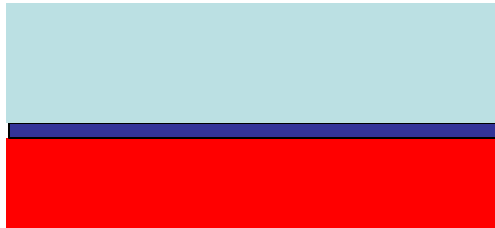
CNM / Substrate 1 (Au, SiN, ...)



Coat with transfer medium



Dissolve substrate 1



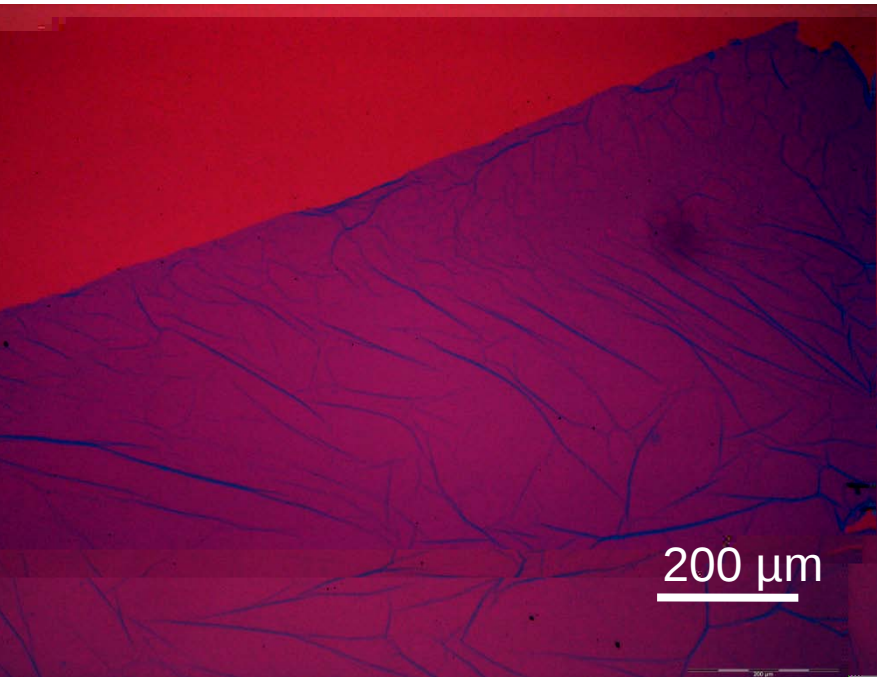
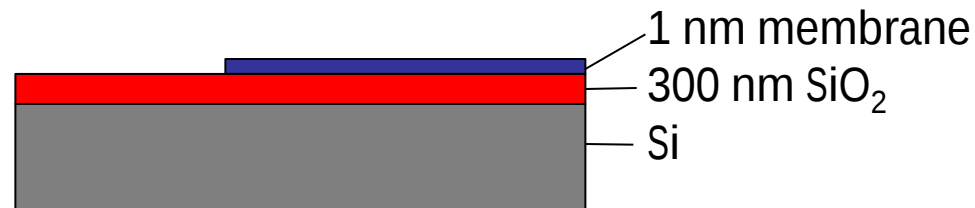
Place on substrate 2



Dissolve transfer medium
CNM / Substrate 2 (SiO, Si, ...)



1 nm thick Membrane on SiO₂/Si: Interference Contrast



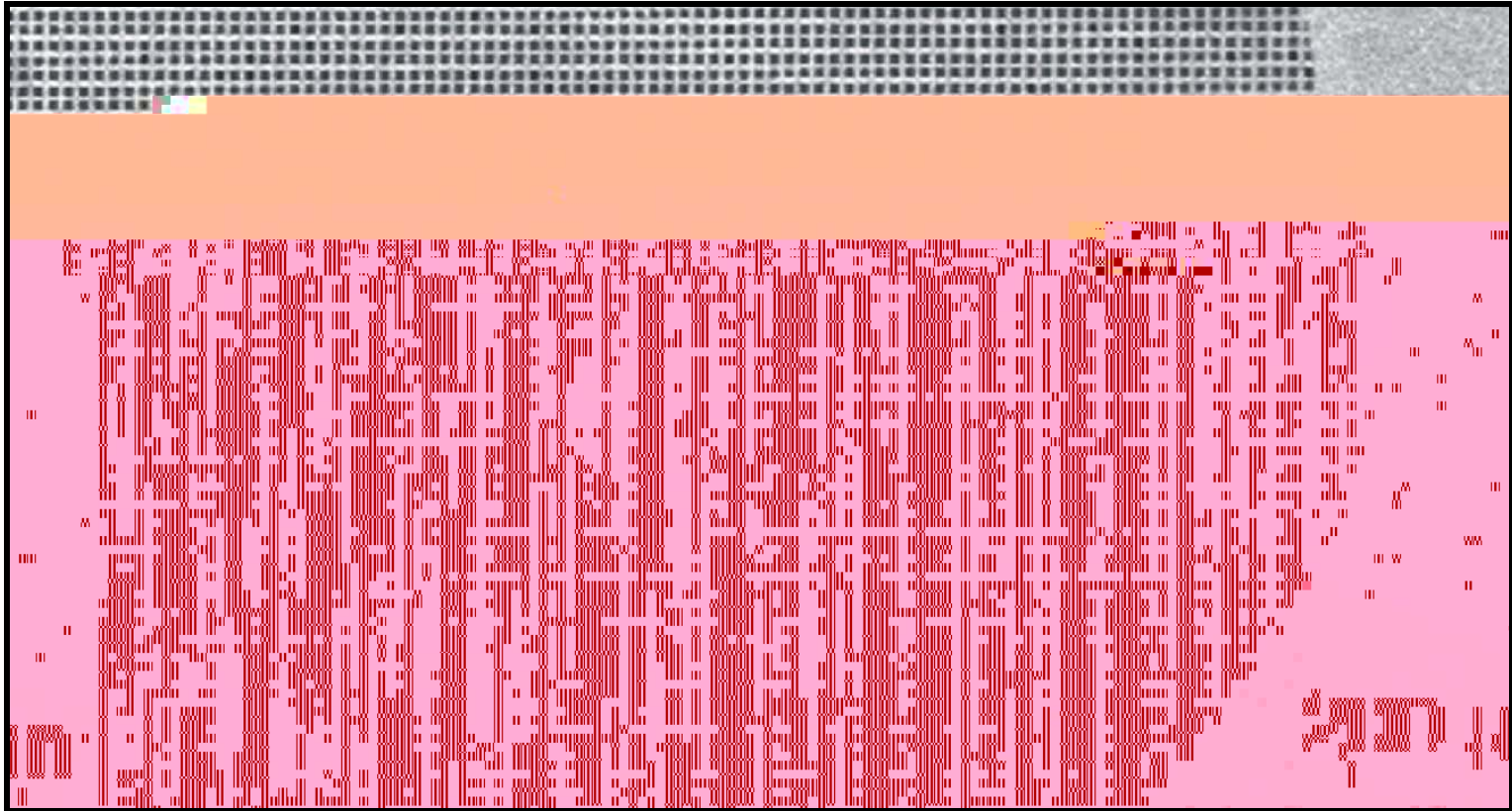
optical micrograph



photograph

Nanosheet on TEM grid

TEM grid (Au 1500mesh), SEM Image

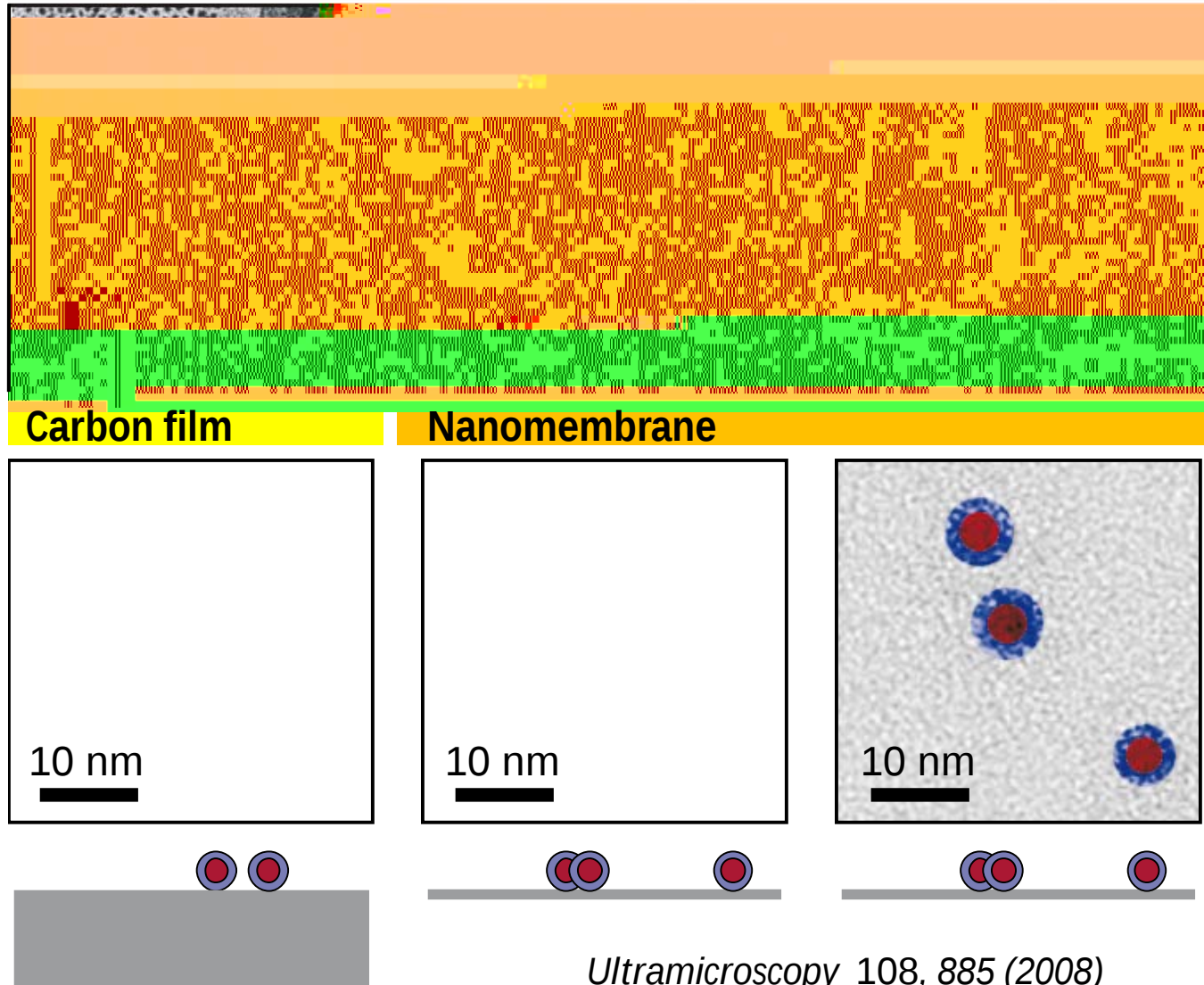


Nottbohm et al., Ultramicroscopy 108, 885 (2008)



100 μm

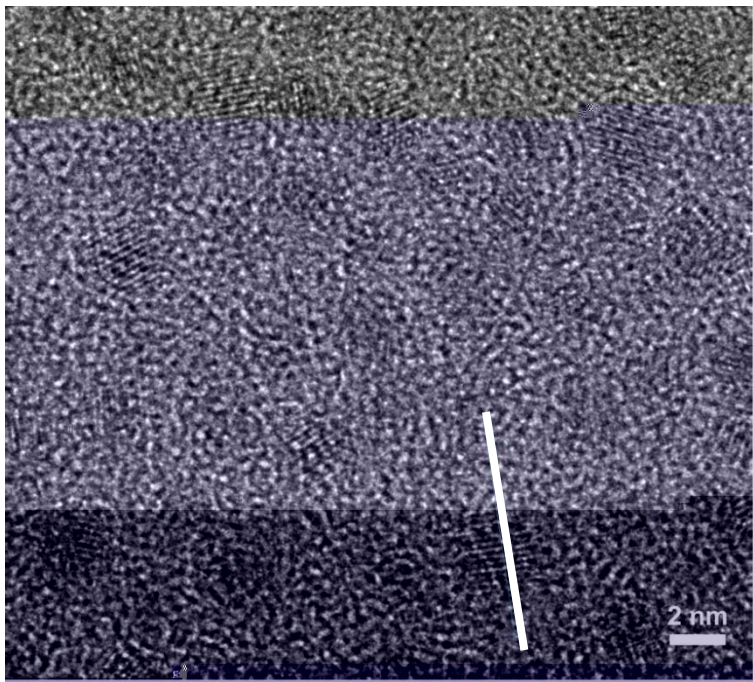
Nanomembrane supports for HRTEM: Imaging of - Co nanoparticles (ca. 4 nm)



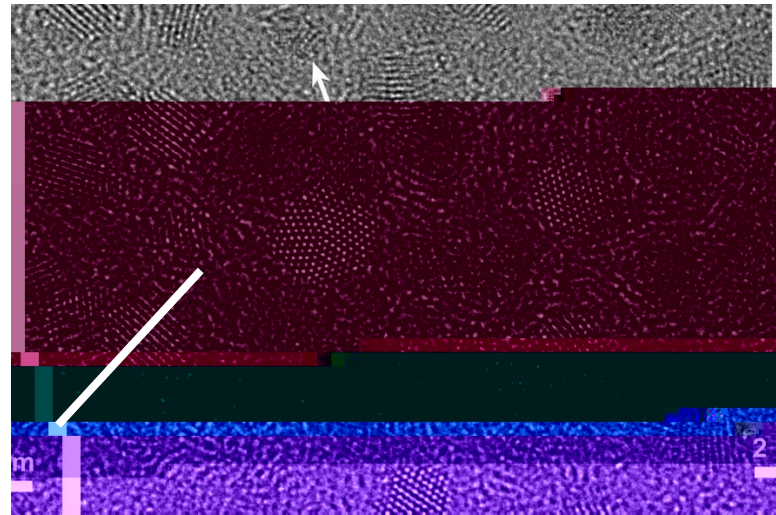
Ultramicroscopy 108, 885 (2008)

TEM - Au₅₅ Cluster

Carbon film



Nanosheet



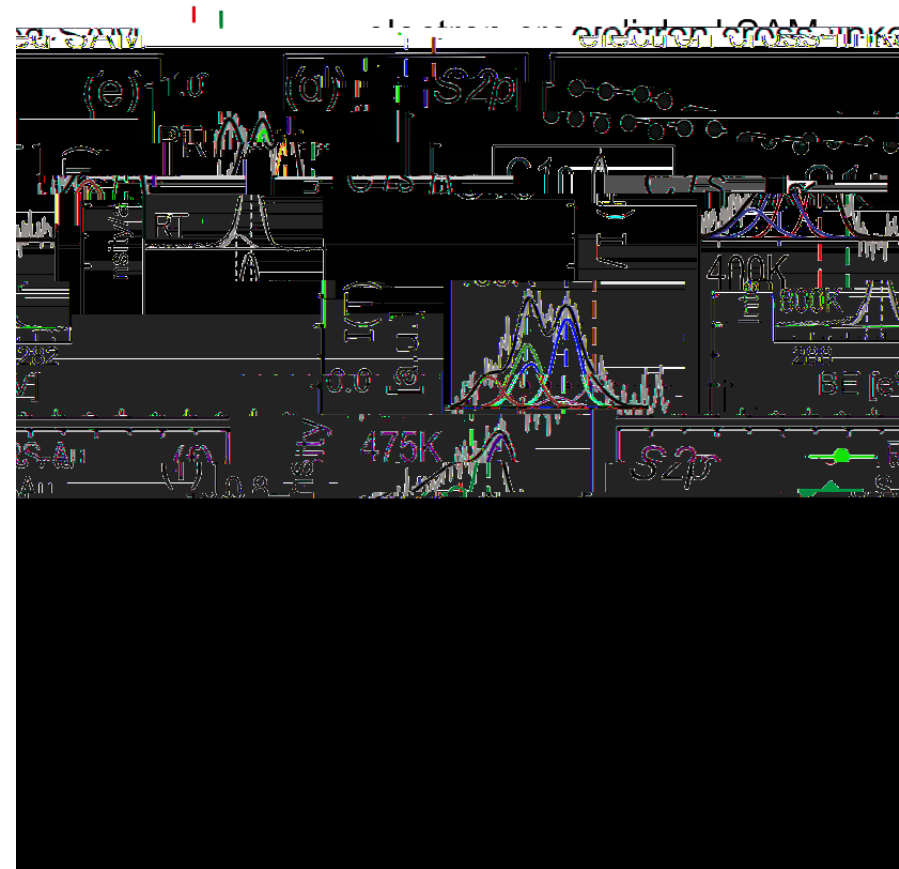
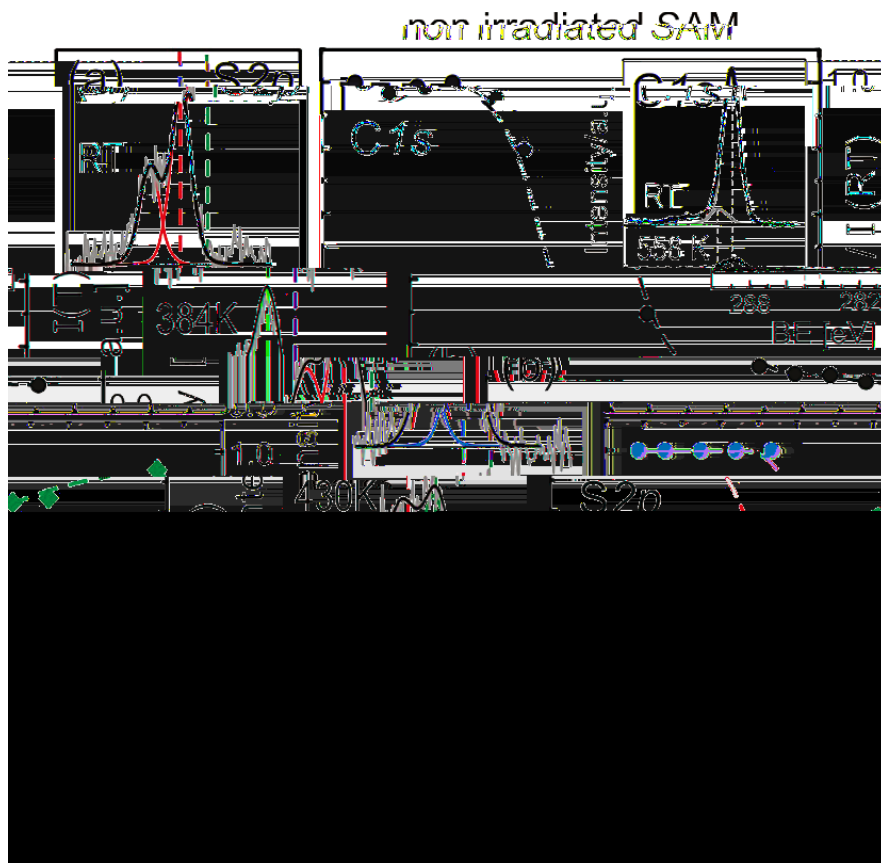
Thermal Properties of Carbon Nanomembranes



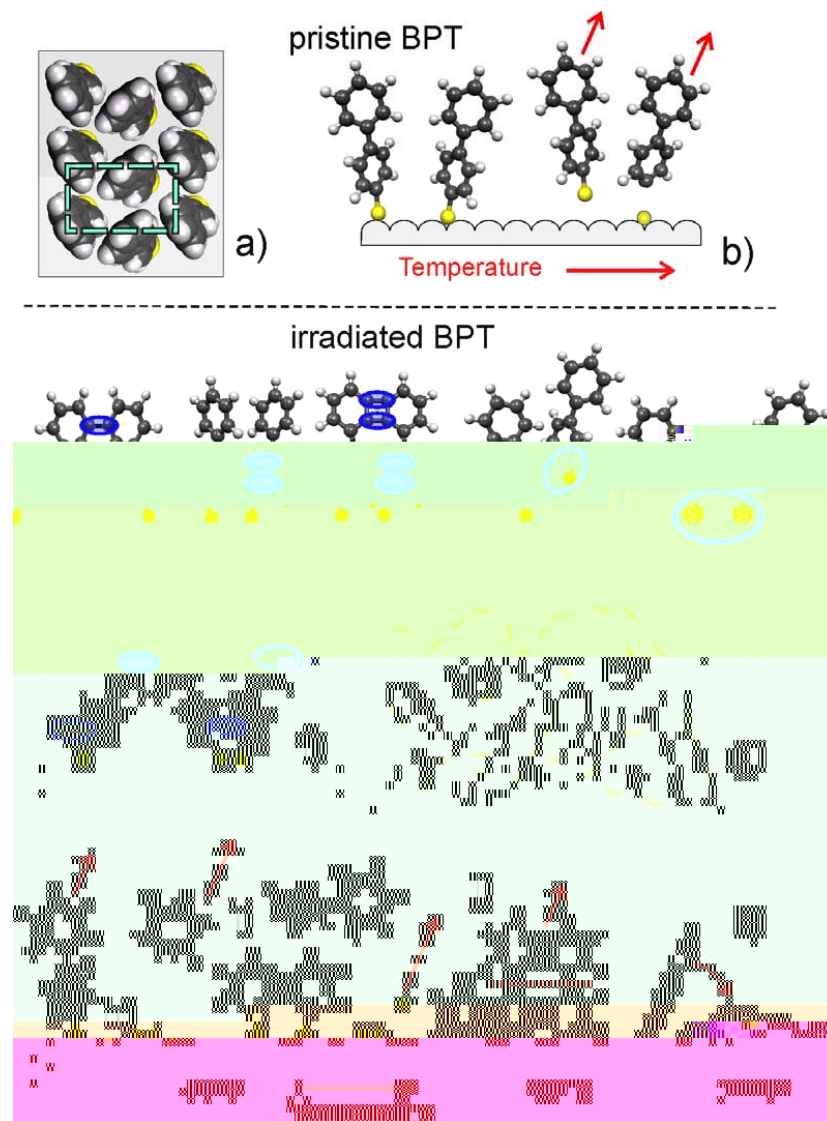
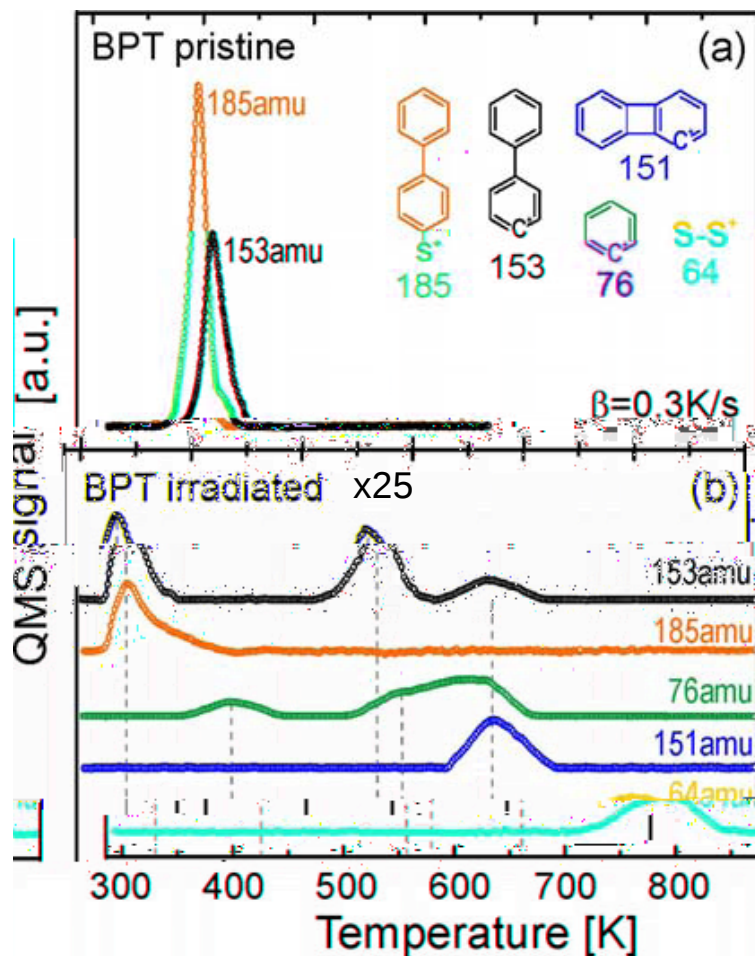
Heating of biphenylthiol nanosheet on SiO₂-Si in UHV

Appl. Phys. Lett. **90**, 053102 (2007)

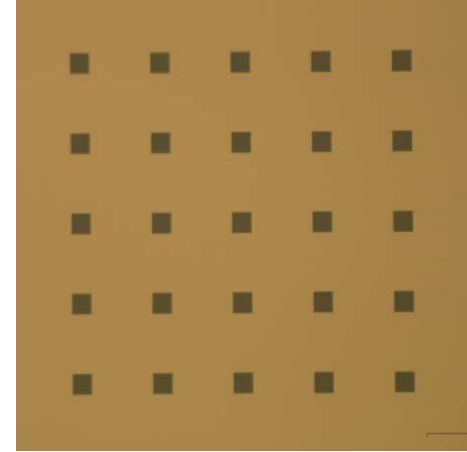
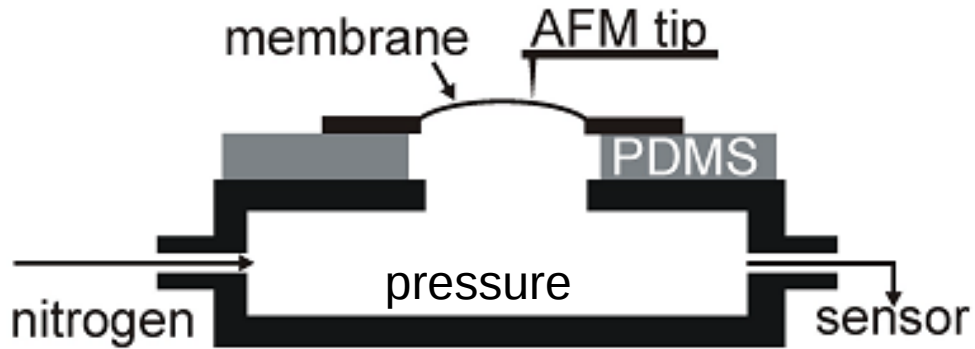
Heating of pristine and cross-linked BPT SAMs (XPS)



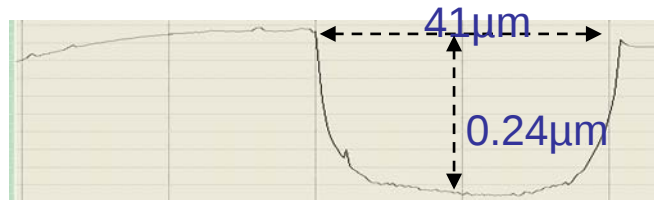
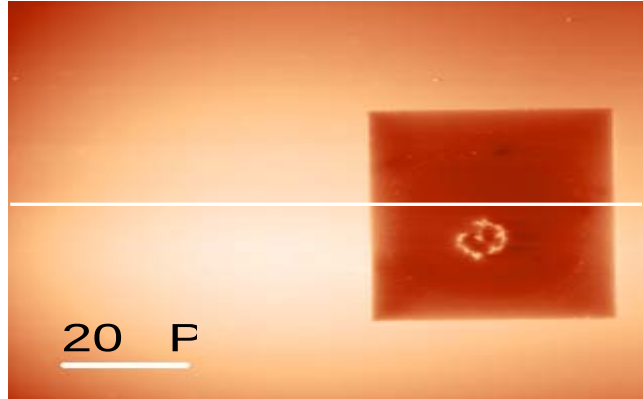
Heat induced molecular desorption



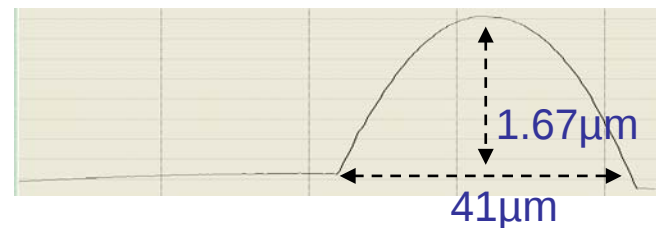
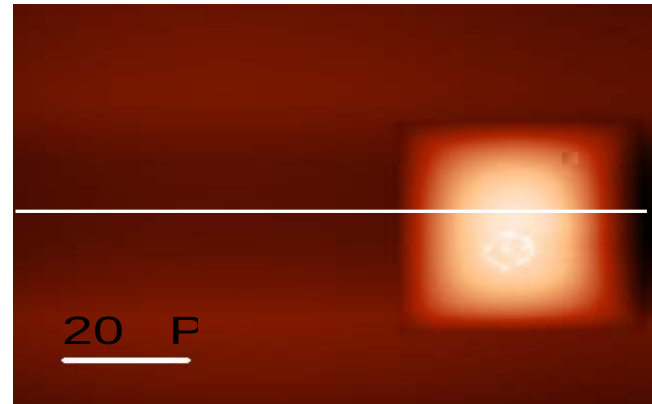
Membrane mechanics: Bulge Tests



$p=0$



$p=750\text{Pa}$



Mechanical properties of nanomembranes

Pressure-deflection relationship for a stressed nanomembrane:

$$P = P_1 + P_2 + \frac{Et}{a^4(1-\nu)}h^3 + \frac{\nu_0 t}{a^2}h$$

J.J.Vlassak and W.D. Nix, J. Mater. Res. 7 (1992) 3242

t=1.5nm: thickness
 $\nu=0.35$: Poisson's ratio
E=Young's modulus
 ν_0 = residual stress
a: half-width
b/a: aspect ratio

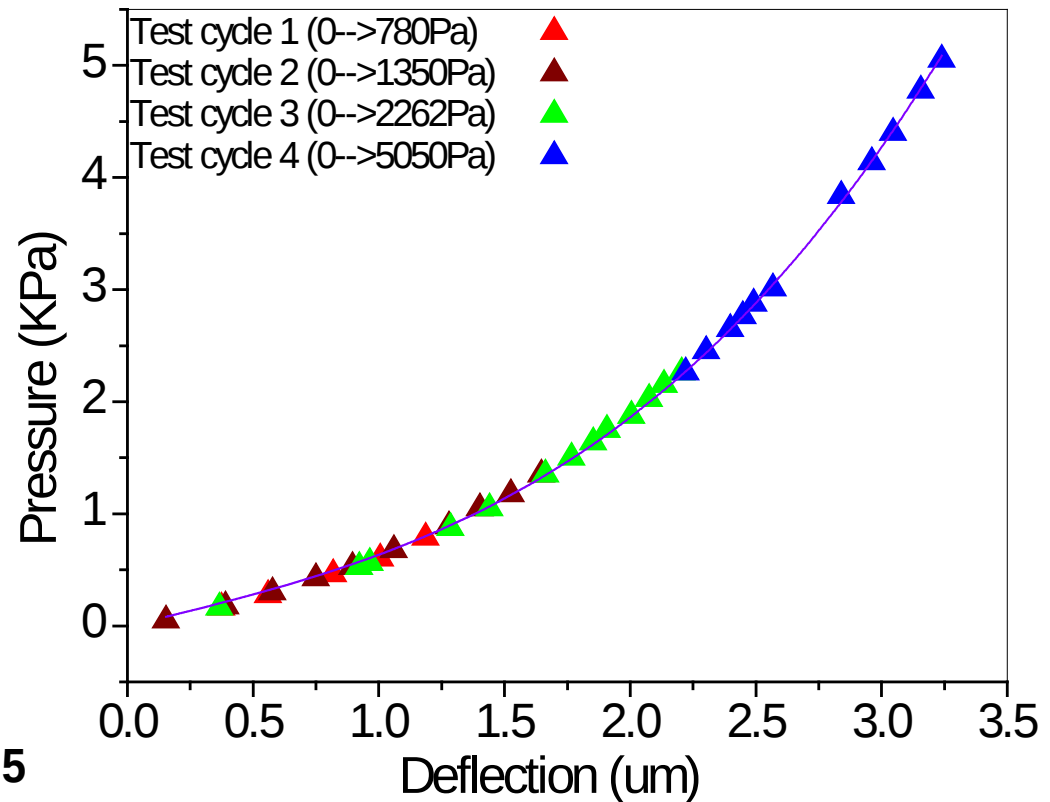
E = 10.0 GPa

ν_0 = 40.0 MPa

some E values (Gpa):

rubber 0.01....0.1, polystyrene 3.0.....3.5

copper 110... 130, diamond 1050...1200



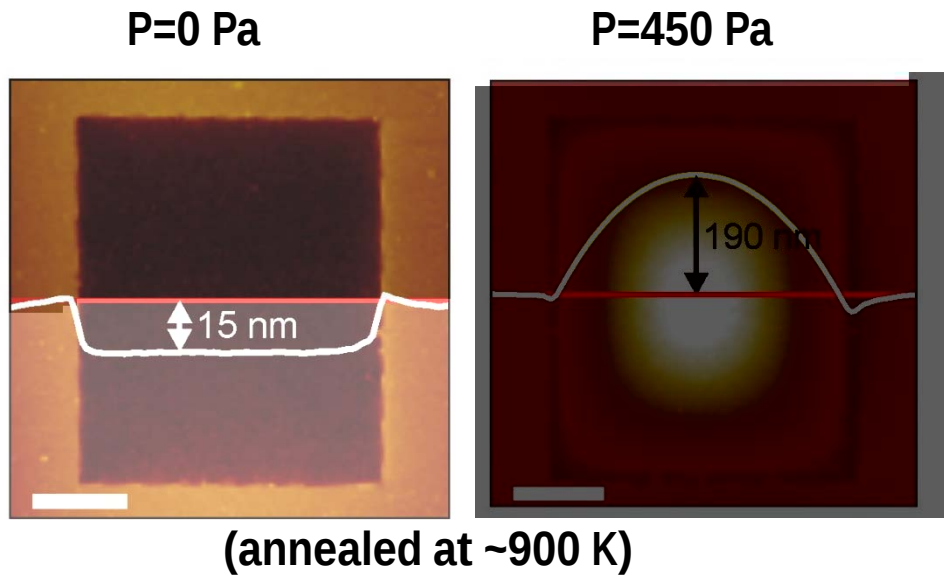
Comparison with other Free-standing Nanomembranes

Freestanding nanomembranes	Thickness (nm)	Fabrication Method	Young's Modulus (GPa)	Tensile Strength (MPa)
Nanocomposite membranes [1]	55	Spin-assisted layer by layer assembly	8± 3.5	40...100
IPNs hybrid nanomembranes[2]	35	Spin-coating and polymerization	N.A.	105
Nanomembranes (epoxy resin)[3]	20	Spin-coating and baking	N.A.	30
Nanomembranes[4] (melamine,phthalic, rethane,epoxy)	19...24	Spin-coating, irradiation, baking	1.2...3.5	10...22
Carbon Nanomembrane	1.5	Self-assembly & cross-linking	10	150...420

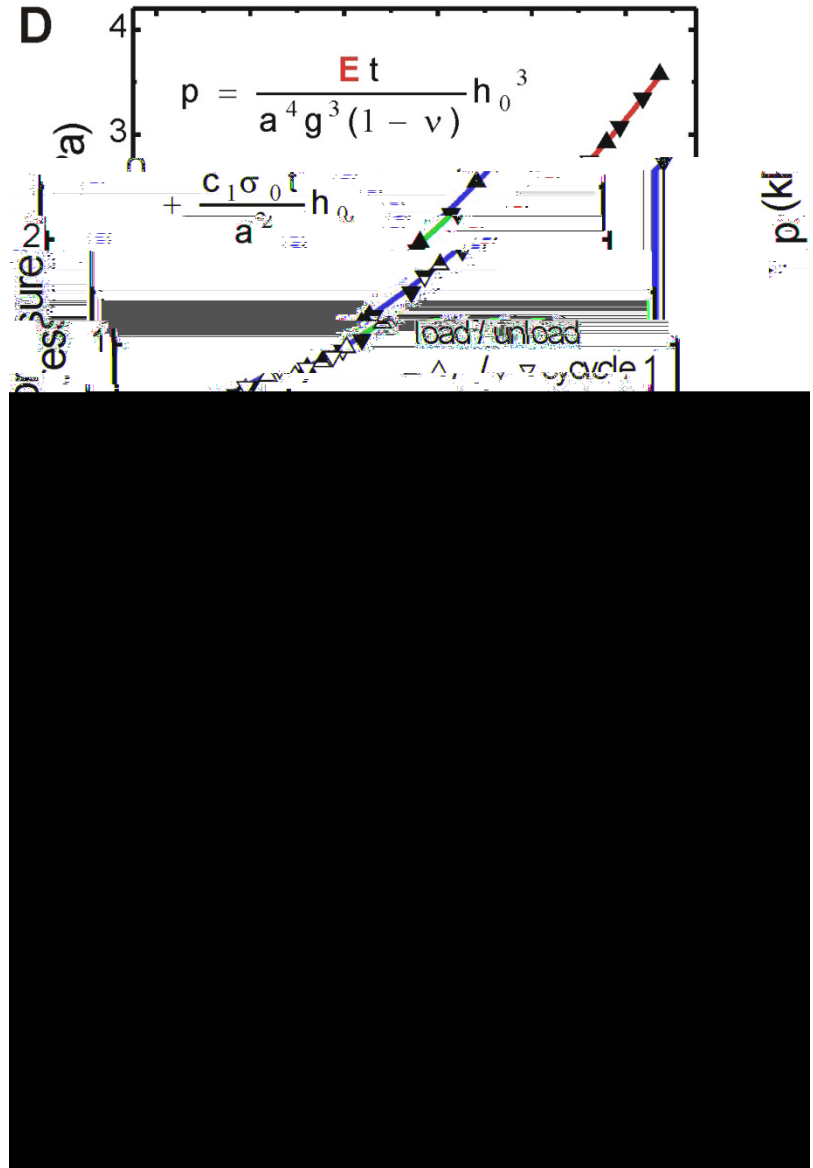
[1] Nature Materials 3 (2004) 721; Advanced Materials 17 (2005) 1669

[2] Nature Materials 5 (2006) 494

Effect of Annealing on Young's Modulus

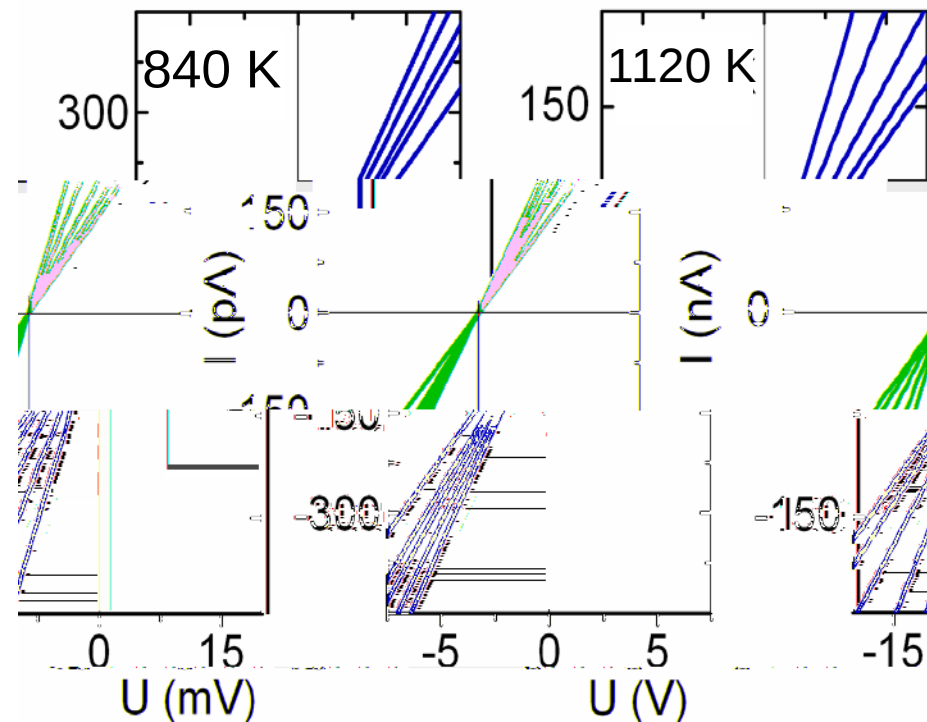
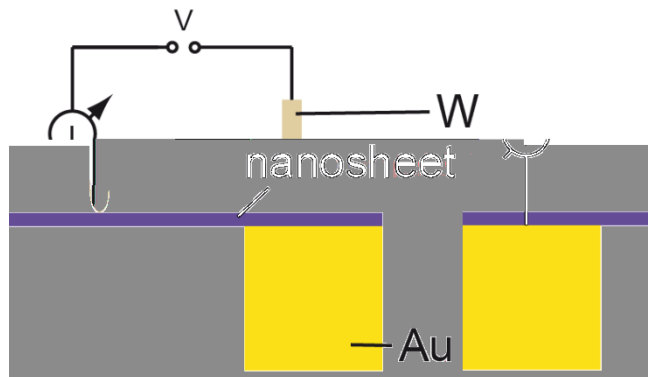


- Young's modulus increase from ~10 GPa to ~45 GPa after heating at ~1000K
- Residual strain reduced from ~0.9 % to ~0.35 %
- Structural transformation



Electrical Characterization of Nanomenbranes:

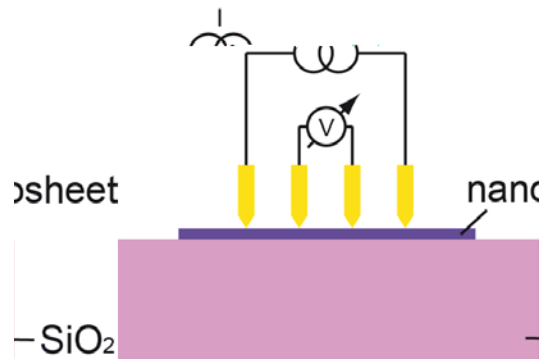
Electrical Characterization of Nanomenbranes: 2-point measurement of free-standing membrane in UHV SEM/STM



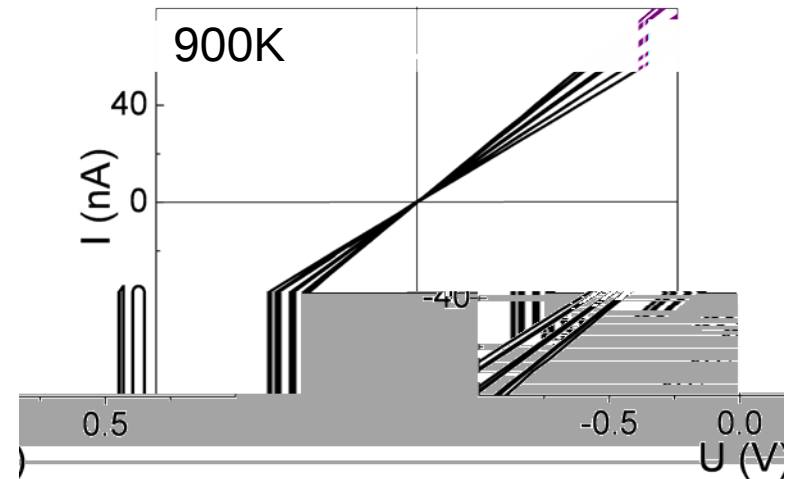
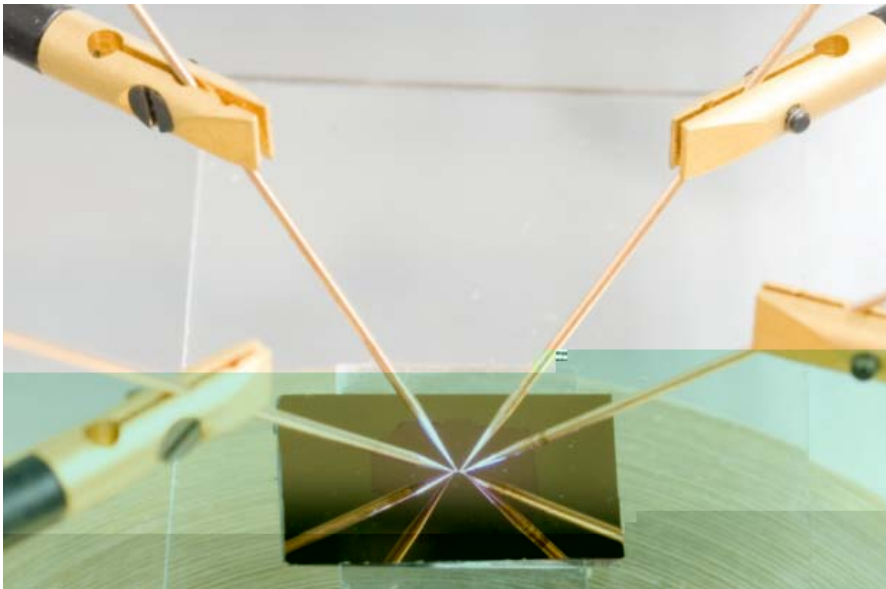
Turchanin et al. Adv. Mater. **21**, 1233 (2009)

Electrical Characterization of Nanomenbranes:

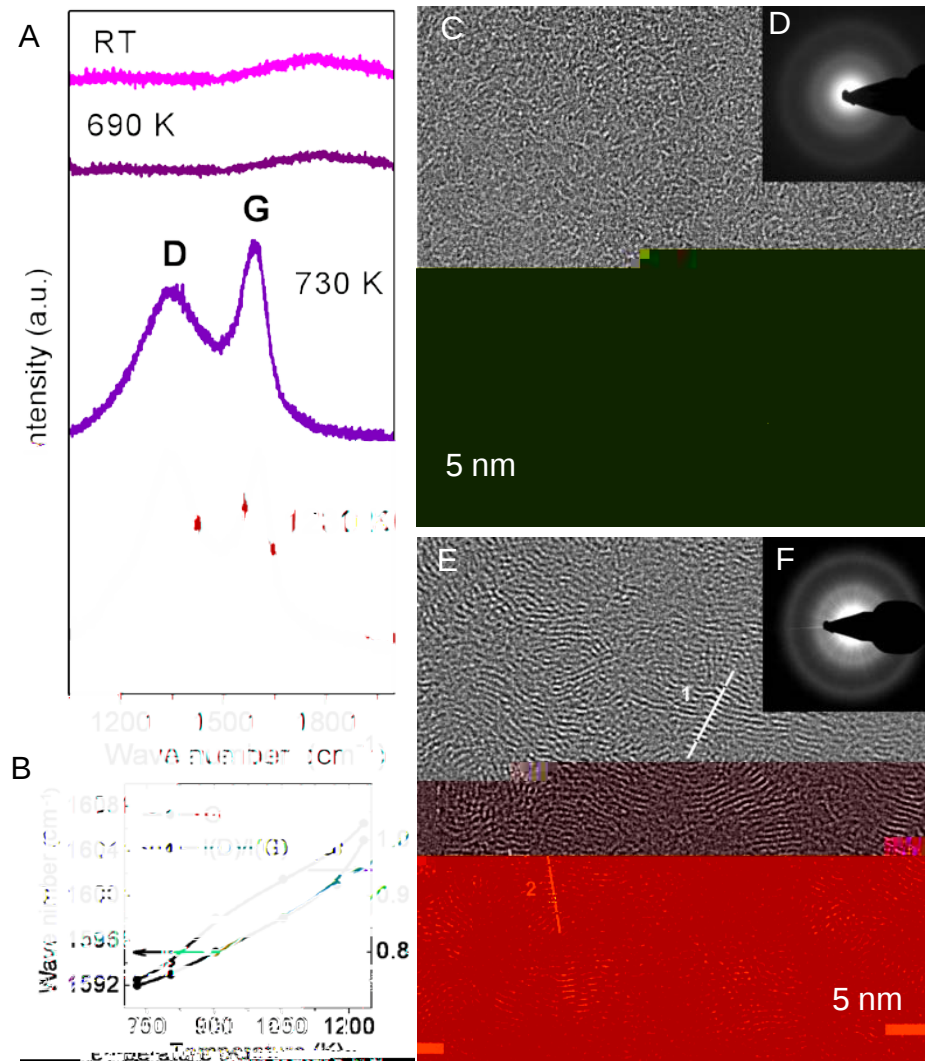
4-point measurement of supported membrane on SiO₂-surface



$$U_s = \frac{S}{\ln(2)} \frac{V}{I} = 4.532 \frac{V}{I} : / \text{square}$$

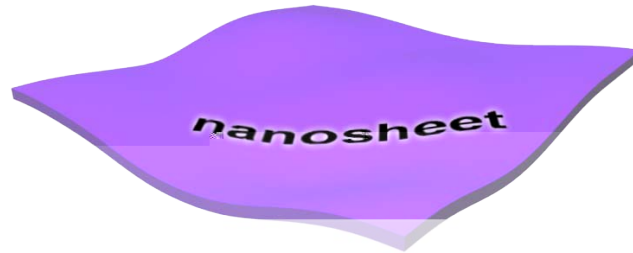


Structural transition (insulator to conductor) in nanomembrane (Raman spectroscopy and TEM):



• Formation of nanocrystalline graphene
 • Tunable electrical properties !!

Perforating and functionalizing carbon nanomembranes

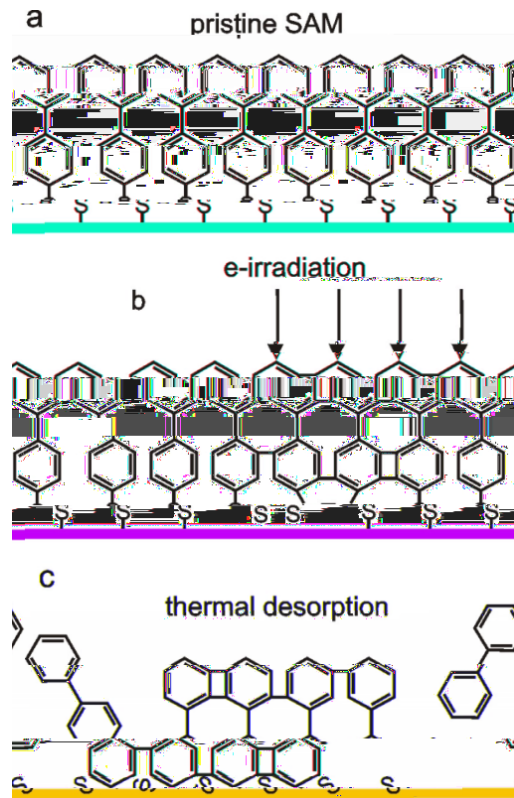


Thermal Desorption Lithography (TDL): Fabrication of Graphenoid Nanoribbons

25

Irradiation dose (mC/cm²)

100

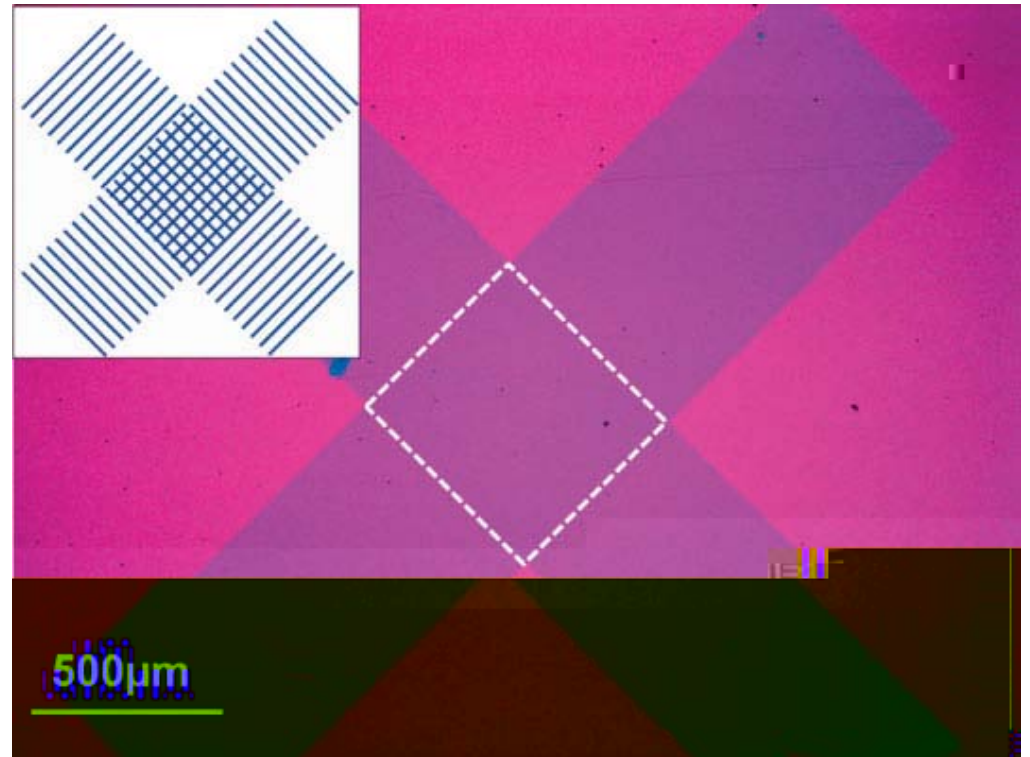
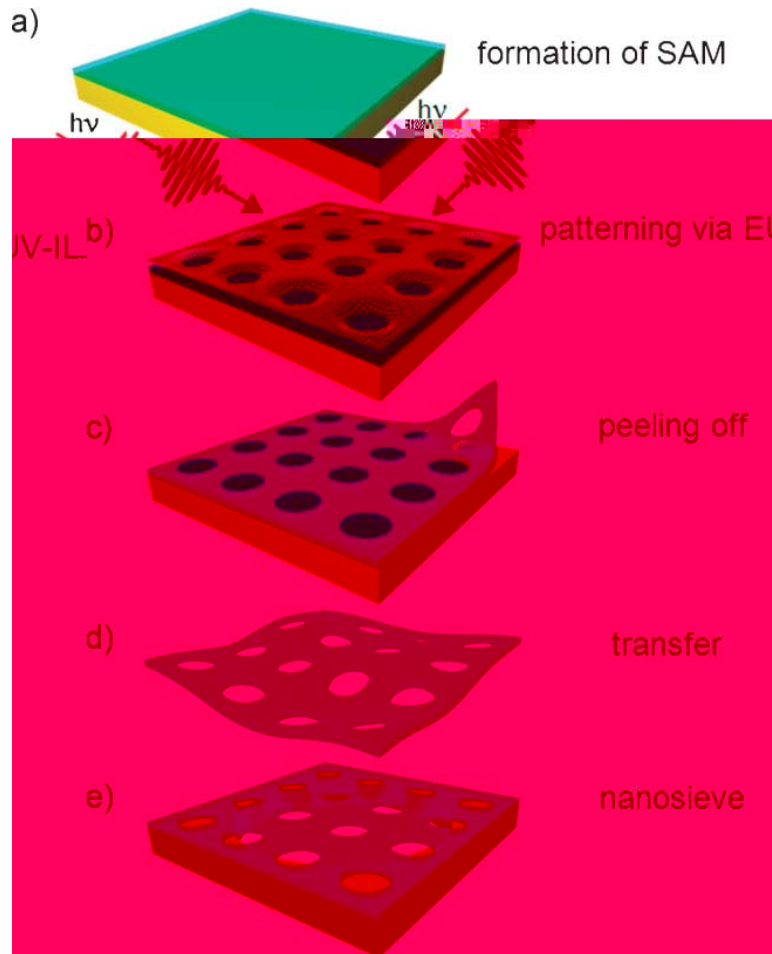


250 Lateral dimension (nm) 10

2μm

Perforated Nanomembranes

Nanosieve fabrication by EUV-IL



Nanosieve membranes with a thickness of 1 nm via EUV-IL

200 x 225 nm
period,

hole diameter =
 138 ± 17 nm

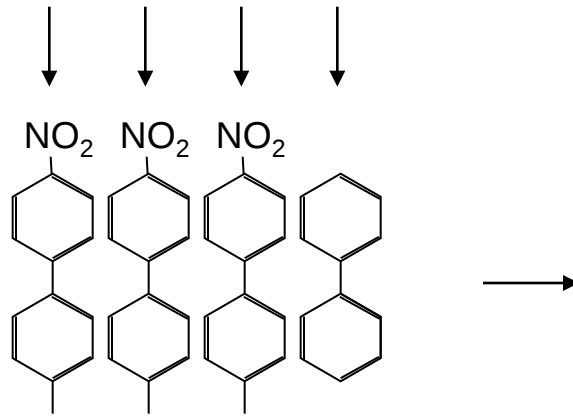
Freestanding nanosieve coated with 5 nm Au



*M. Schnietz et al.,
Small 23, 2651 (2009)*

Electron induced cross-linking and NO_2 to NH_2 conversion “Chemical Lithography”

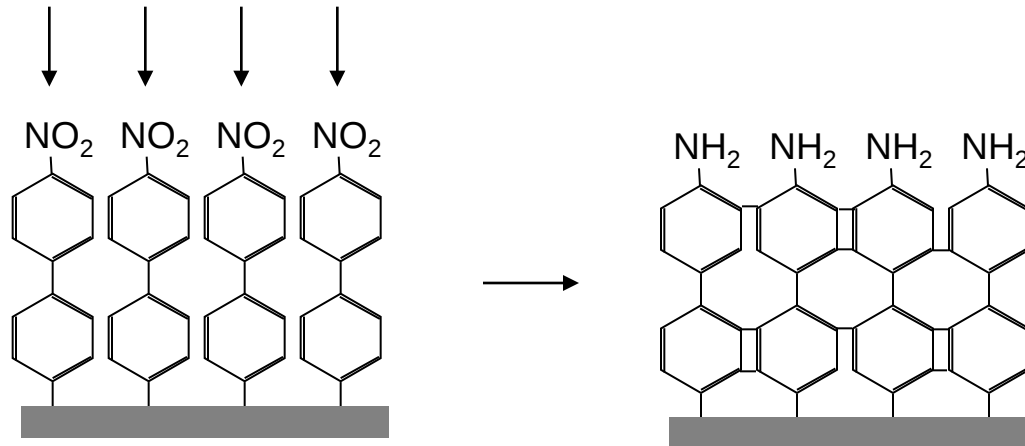
Electrons, 10 -500 eV
area dose: 1 -10 mC / cm^2



Appl. Phys. Lett. 75, 2401 (1999)

Electron induced cross-linking and NO_2 to NH_2 conversion “Chemical Lithography”

Electrons, 10 -500 eV
area dose: 1 -10 mC / cm^2

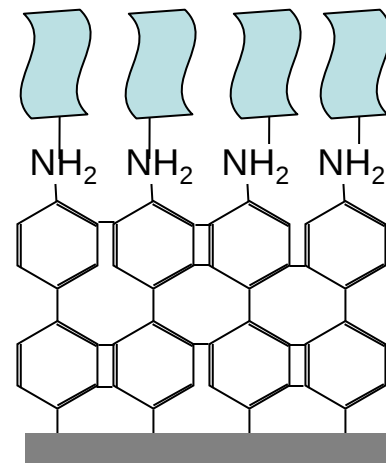
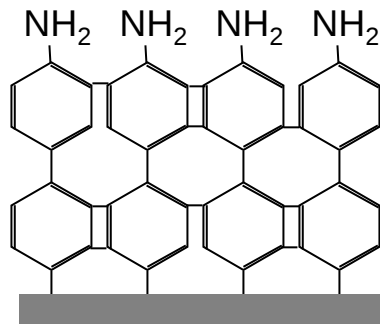
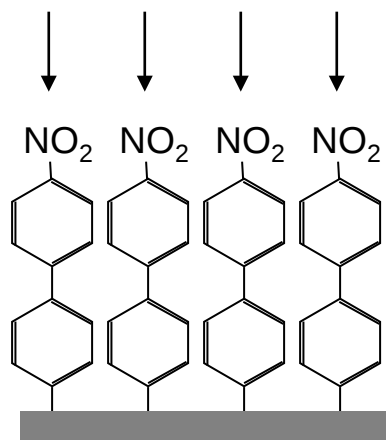


Appl. Phys. Lett. 75, 2401 (1999)

Adv. Mater. 12, 805 (2000)

Electron induced cross-linking and NO_2 to NH_2 conversion: Chemical lithography and subsequent functionalisation

Electrons, 10 -500 eV
area dose: 1 -10 mC / cm^2



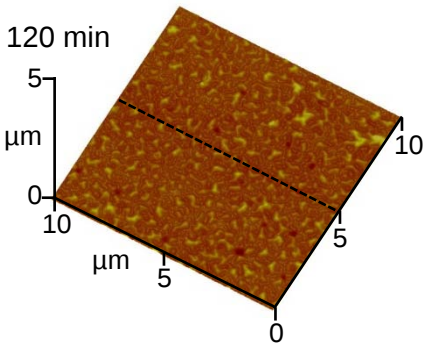
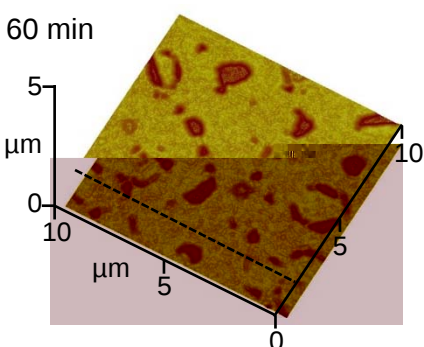
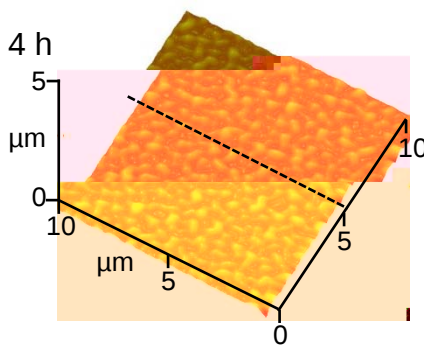
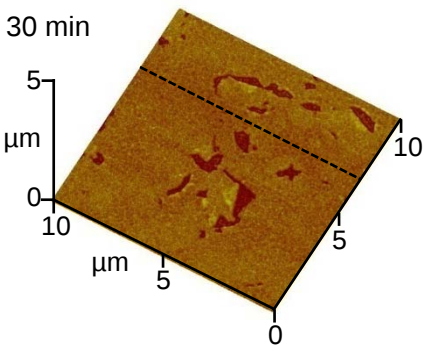
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Molecules binding to NH_2 group

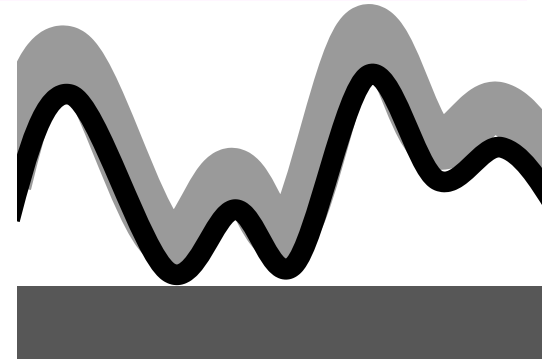
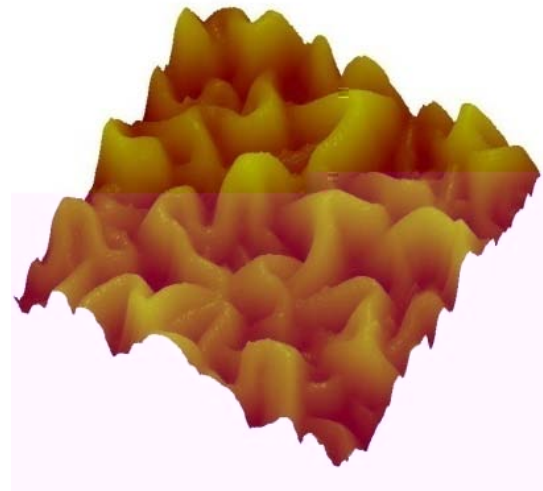
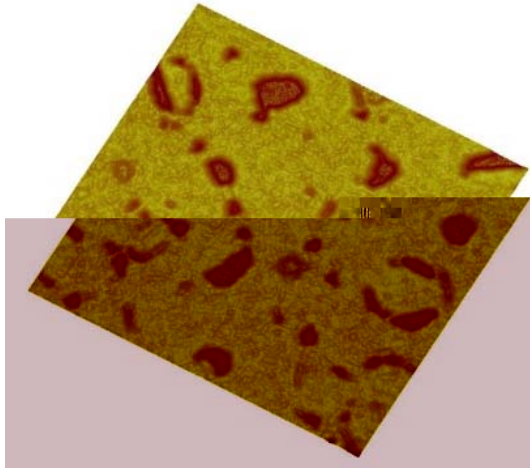
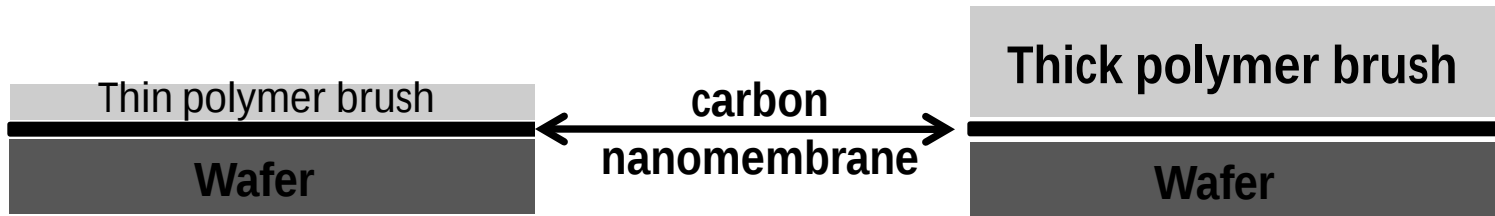
- polymers
- proteins
- dyes
- ...

Polymer Carpets



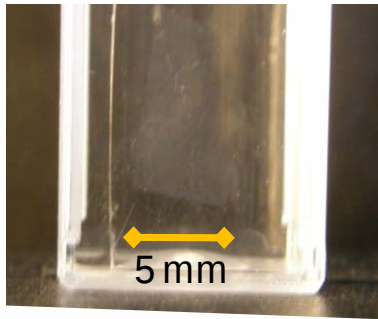


Buckling of Polymer Carpets

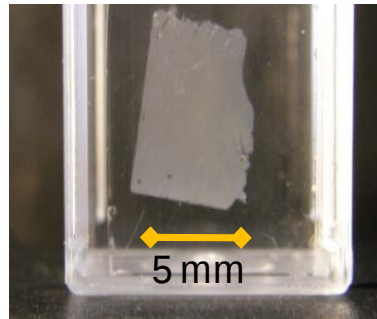


Building a Sensor (and Actuator) by Buckling of Polymer Carpet

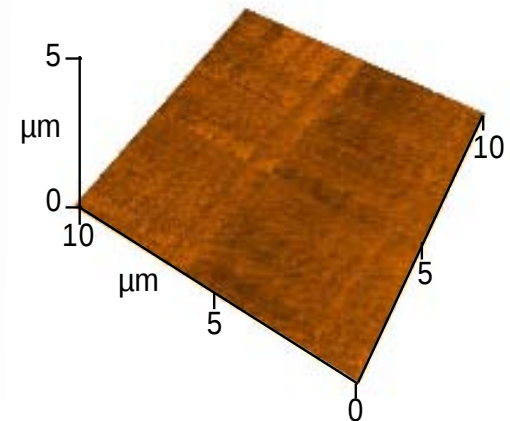
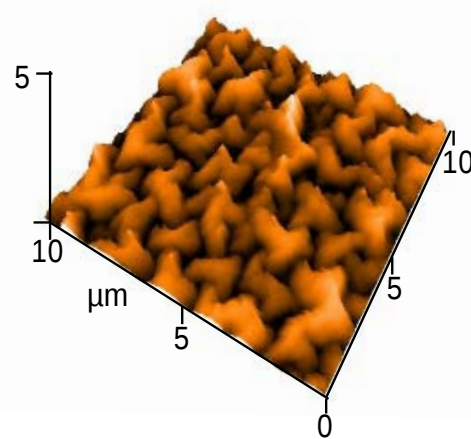
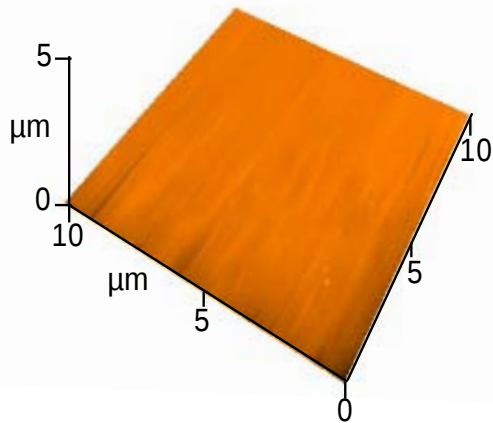
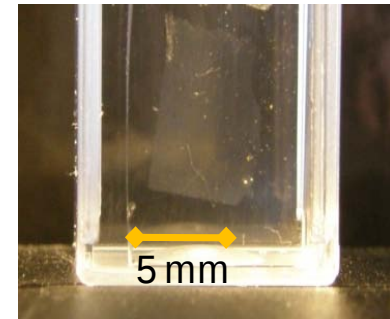
a) EtOH



b) H₂O pH = 7



c) H₂O pH = 2.5



Photographs and AFM measurements of P4VP carpets in
a) ethanol, b) water at pH 7 and c) water at pH 2.5.

Electron Transmission through freestanding Polymer Carpets

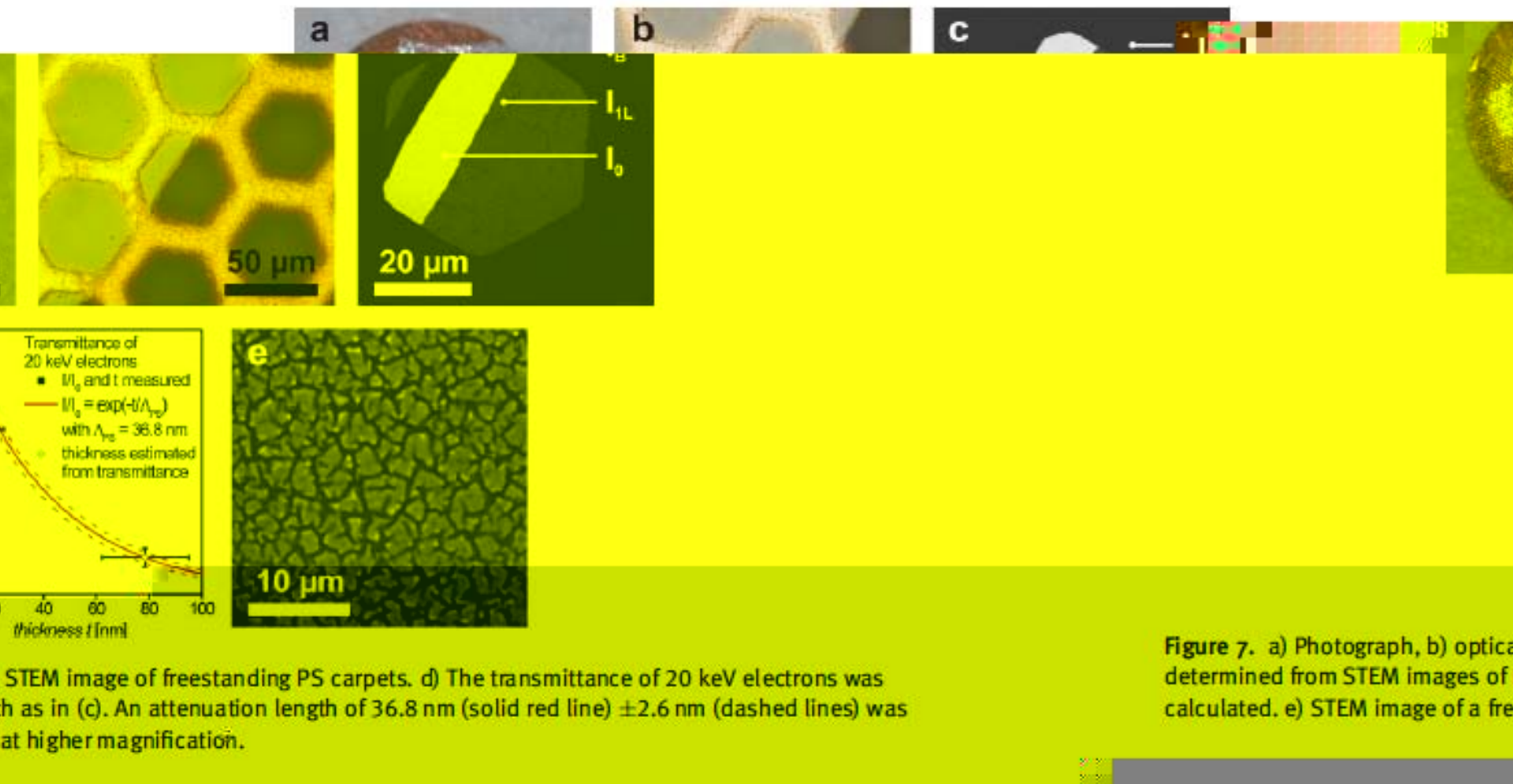
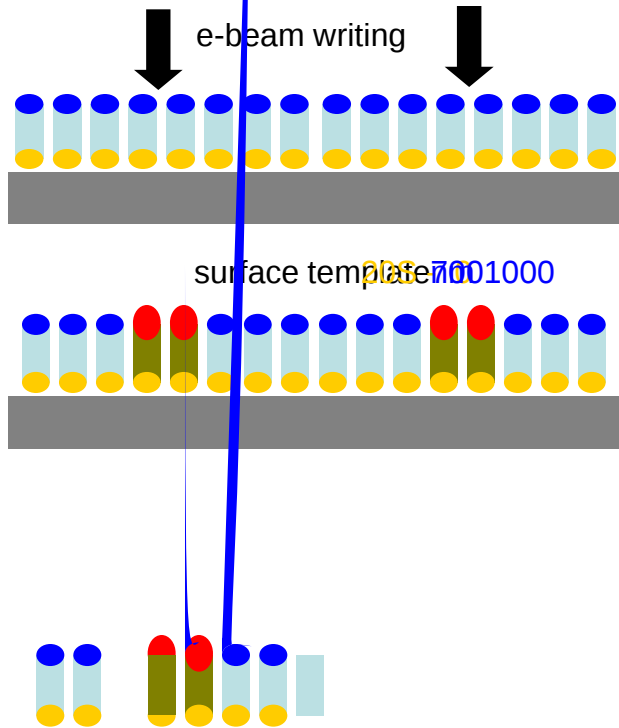


Figure 7. a) Photograph, b) optical micrograph, c) photograph, d) plot of transmittance of 20 keV electrons, e) STEM image of a freestanding PS carpet.

STEM image of freestanding PS carpets. d) The transmittance of 20 keV electrons was determined from STEM images of freestanding PS carpets. An attenuation length of 36.8 nm (solid red line) ± 2.6 nm (dashed lines) was determined from the transmittance data at higher magnification.

Immobilization of biomolecules

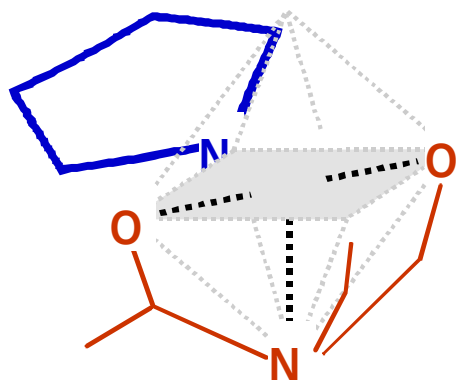


NTA/His-tag interaction: molecular tweezers

histidine

histidine tag (His-tag)

{easily introduced into
proteins by genetic engineering



NTA

nitrilotriacetic acid (NTA)

{high specificity for neighboring
histidine residues

{easily functionalized

reversible binding

imidazole

ETDA

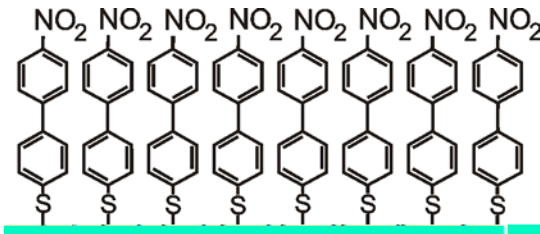
low pH

high affinity by utilization of

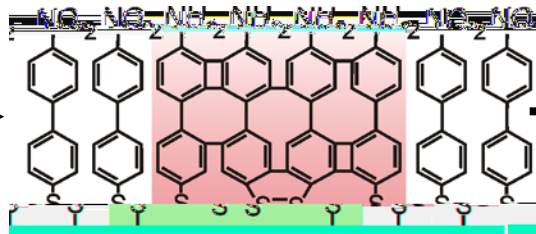
multivalent chelators: bis-NTA, tris-NTA

Assembly of the structured chips: schematic representation

SAM formation



e-beam/EUV lithography

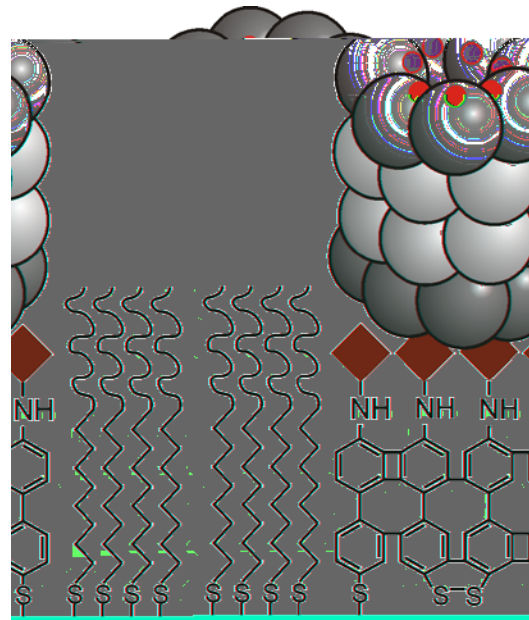
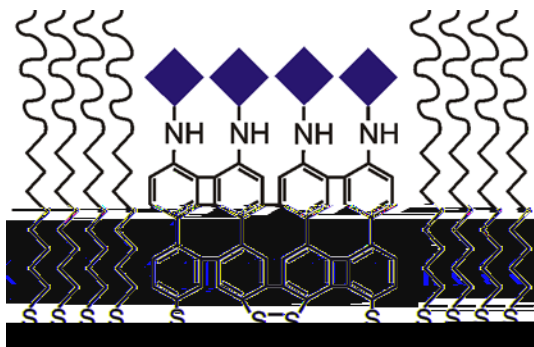


molecular self-assembly
e-beam/EUV lithography
chemical biology
molecular recognition

functional immobilization of His₆-tagged proteins

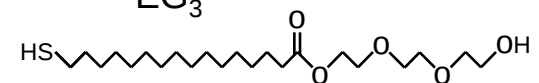
tris-NTA

grafting of multivalent chelators
and generation of the protein
repellent matrix

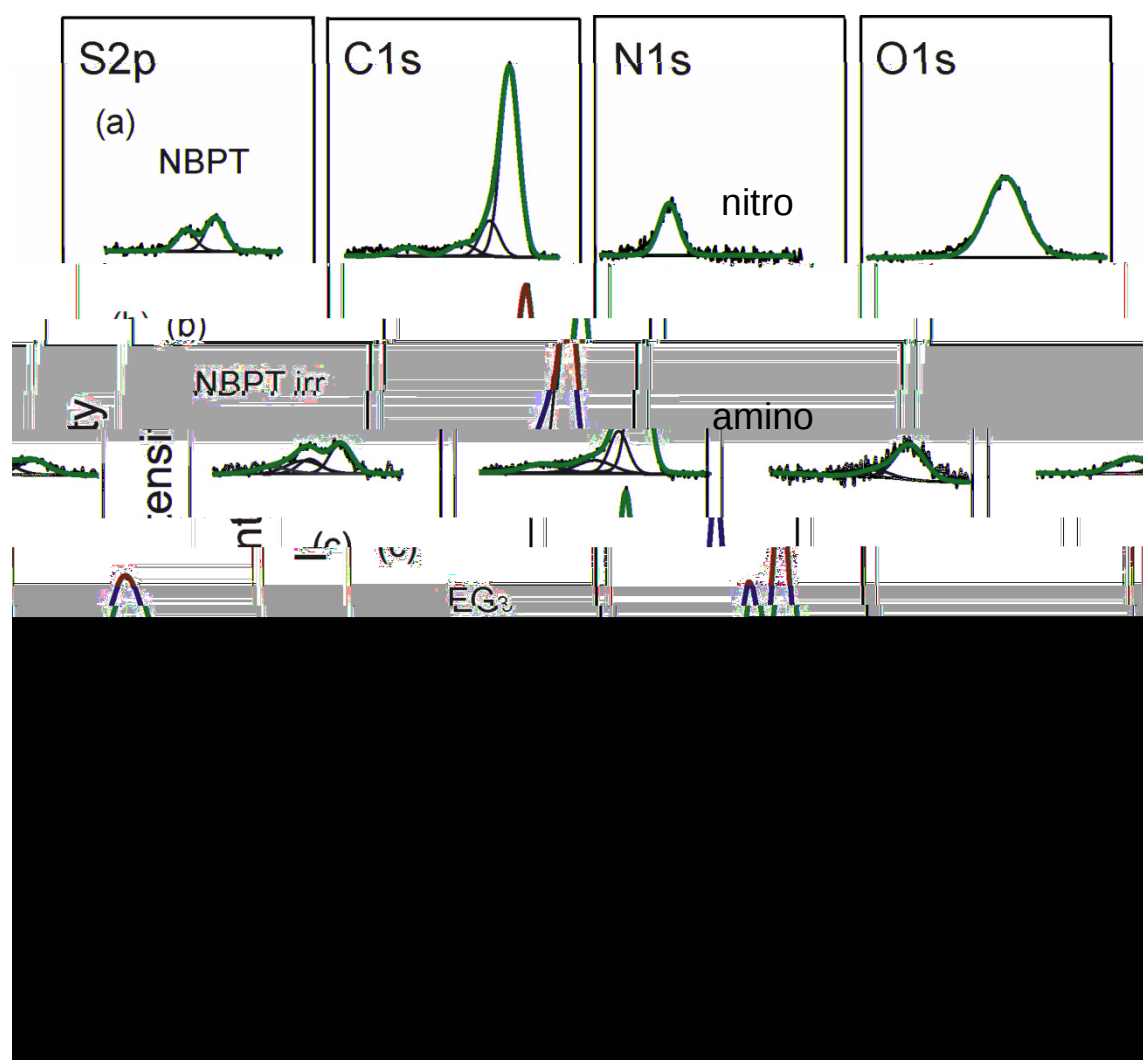


0

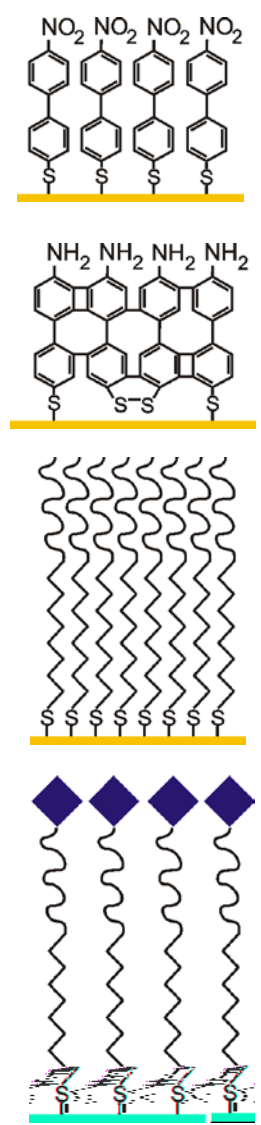
EG₃



XPS characterization of the elemental components of the chip' surface

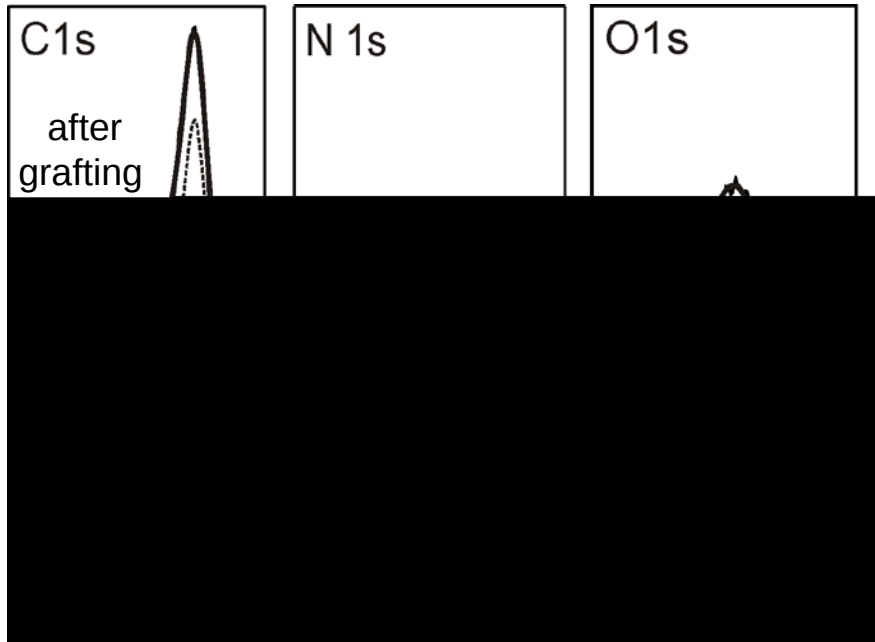


different types SAMs



A.Turchanin, A. Tinazli , M. El-Desawy , H. Großmann, M. Schnietz ,
H. H. Solak, R. Tampé, A. Götzhäuser, Adv. Mater. 20, 471 (2008)

Grafting of multivalent chelators (tris-NTA)



thickness increase $\sim 6 \text{ \AA}$

tris -NTA:NBPT $\sim 1:9$

experimental C:O:N = 11.2:3.5:1

theoretical C:O:N = 11.1:3.4:1

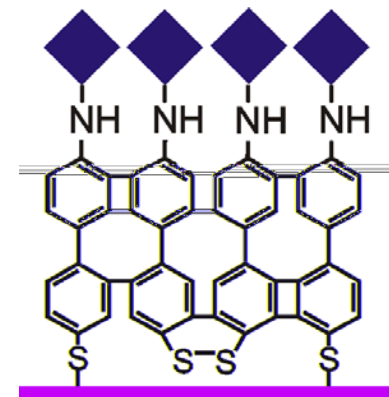
C1s^I 284.9 eV (alkane-like groups)

C1s^{II} 286.8 eV (N-C bonds)

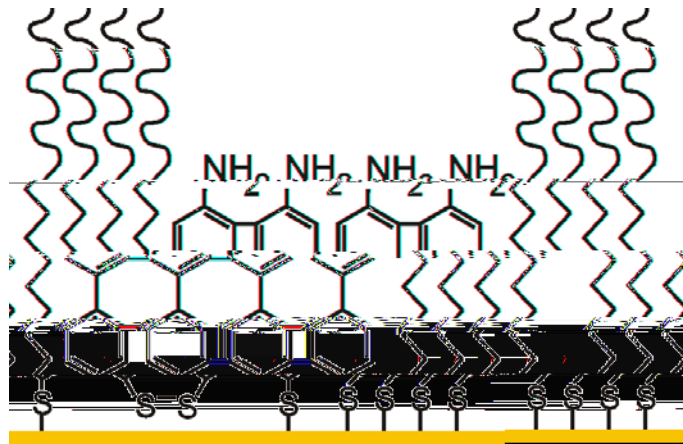
C1s^{III} 288.9 eV (carboxylic groups)

N1s 399.8 eV (amine groups)

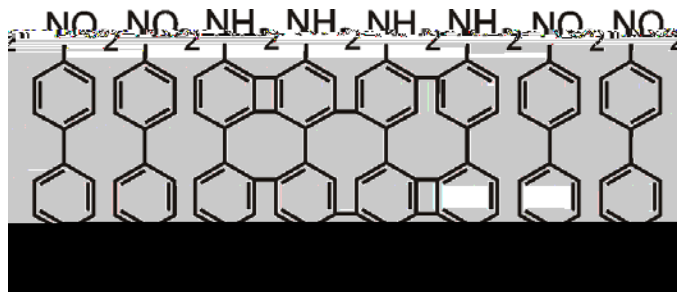
O1s 531.9 eV (carboxylic groups)



Generation of protein repellent matrix by exchange



immersion in ethylene glycol (EG_3)



pristine
SAM

irradiated
SAM

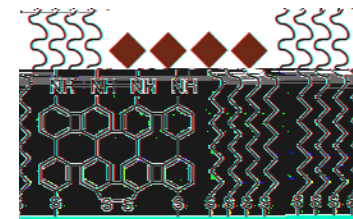
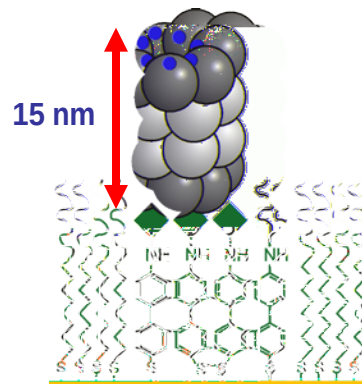
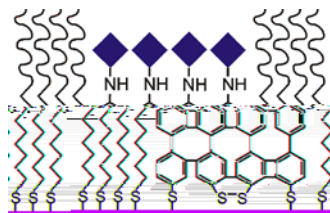
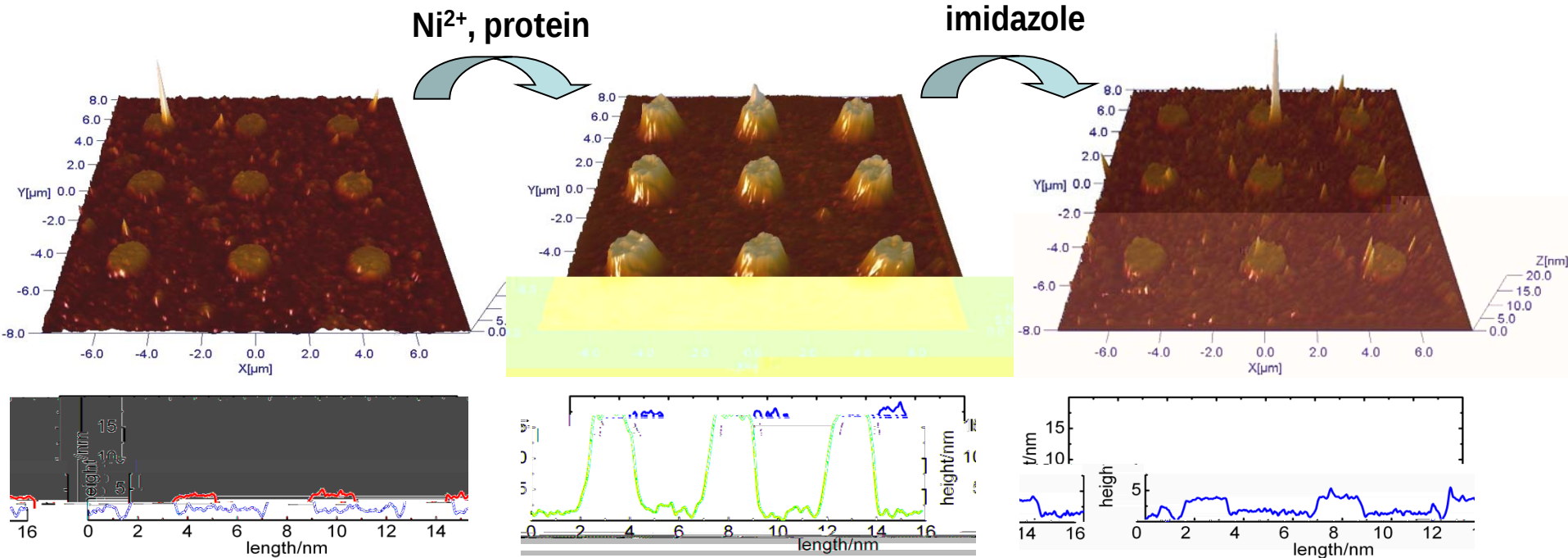
pristine
SAM

Protein chip functioning: an in situ AFM study

Chips' surface in buffer

Immobilization of proteins
(20S His₆-tagged proteasome)

Regenerated chip

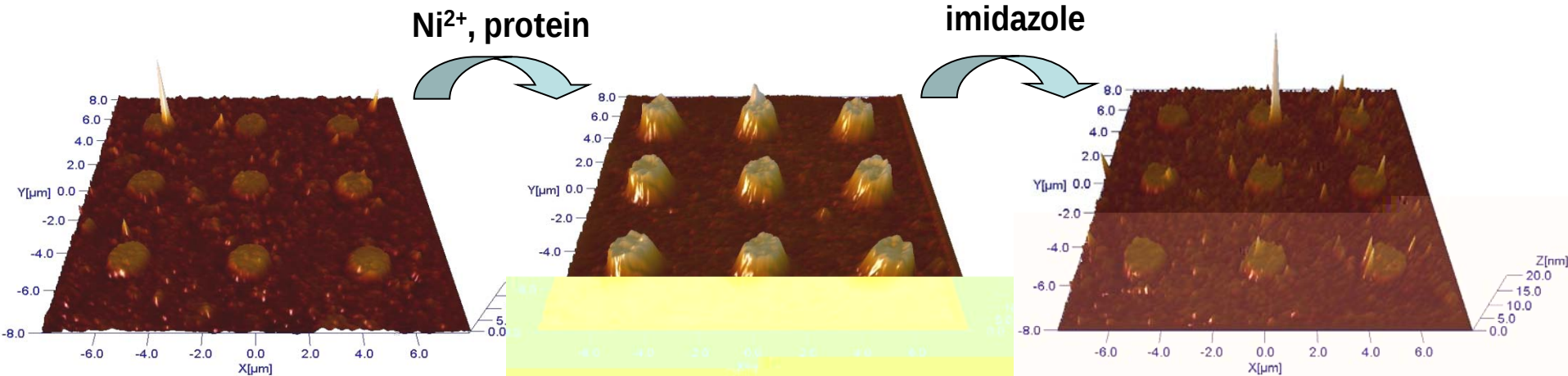


Protein chip functioning: an in situ AFM study

Chips' surface in buffer

Immobilization of proteins
(20S His₆-tagged proteasome)

Regenerated chip



structured
specific
highly parallel
highly affine
oriented
reversible

immobilization of protein micro arrays

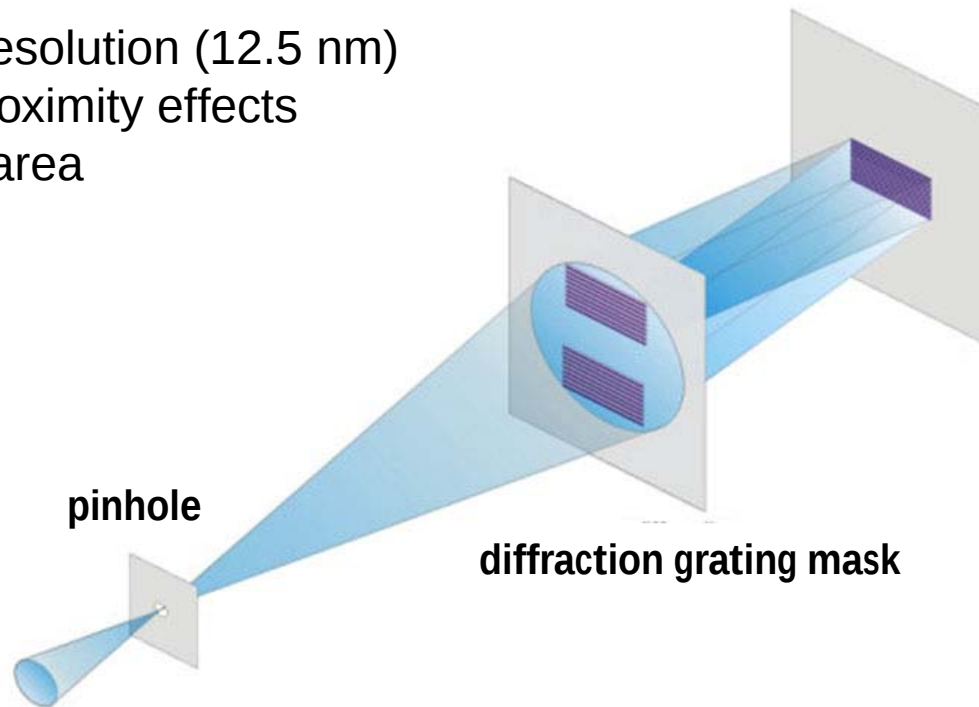
Down to the single molecular resolution?

A.Turchanin et al.
Adv. Mater. 20, 471 (2008)

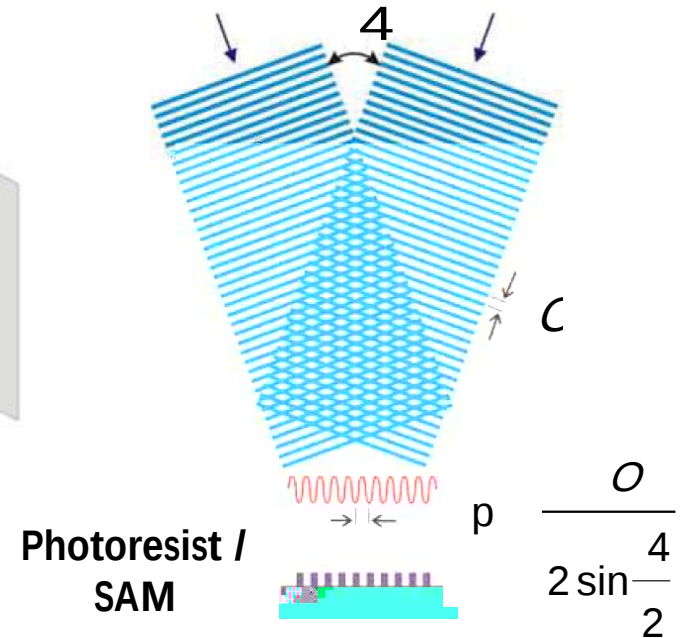
Protein nanopatterns by EUV Interference Lithography

Extreme UV Interference lithography (EUV-IL):

- high resolution (12.5 nm)
- low proximity effects
- large area



**focused synchrotron irradiation
(92.5 eV, 13.5 nm)**

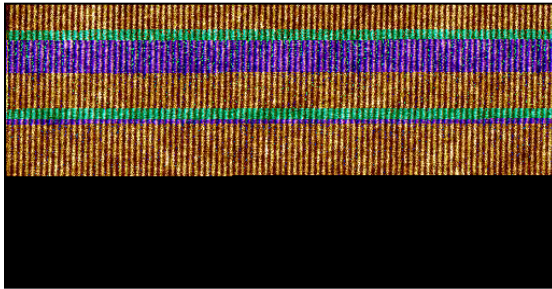


Two coherent beams are forming a linear fringe pattern with a sinusoidal intensity distribution.

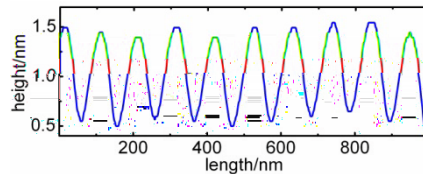
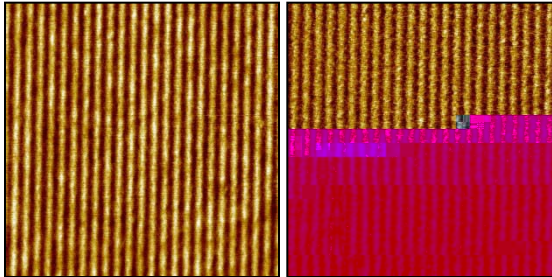
High resolution chemical patterns by EUV-IL: AFM

50 nm lines

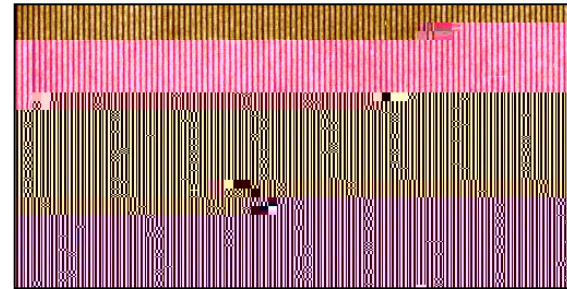
(a) nitro/amino lines, topography, $10 \mu\text{m}^2$



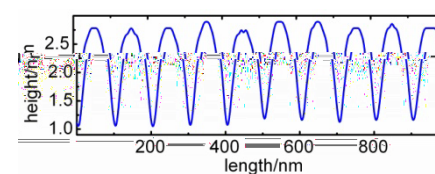
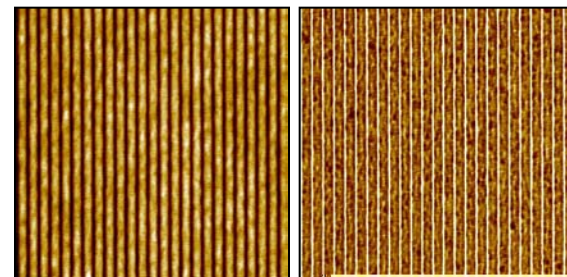
topography (left) and phase contrast (right), $2.5 \mu\text{m}^2$



(b) EG₃-OH /amino lines, topography, $10 \mu\text{m}^2$



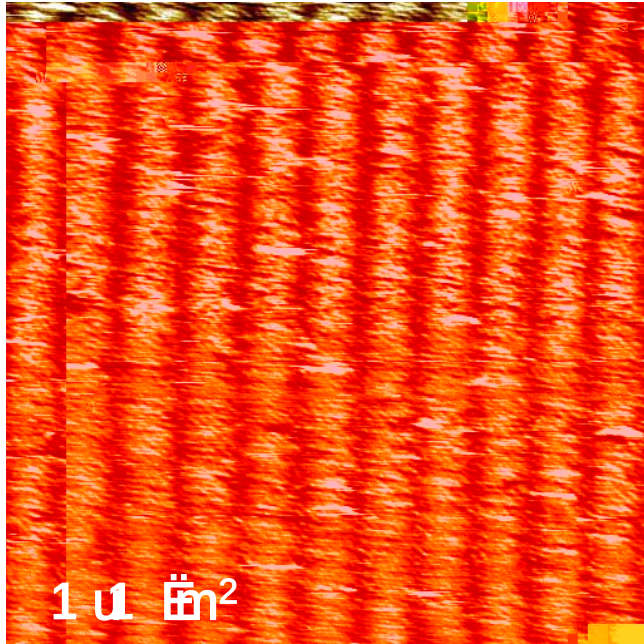
topography (left) and phase contrast (right), $2.5 \mu\text{m}^2$



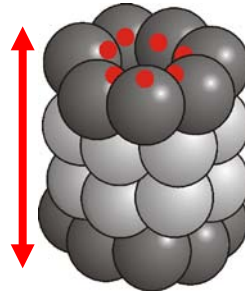
Immobilization of protein nanoarrays : in situ AFM characterization

Proteasome lines

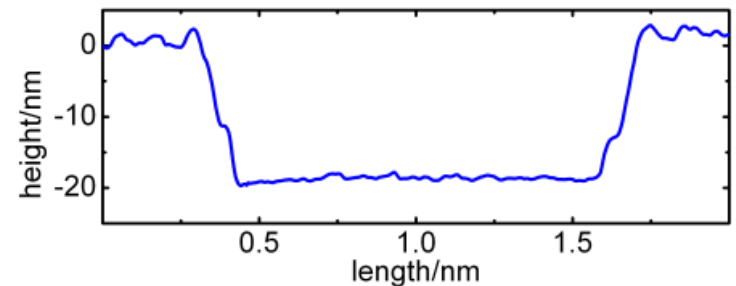
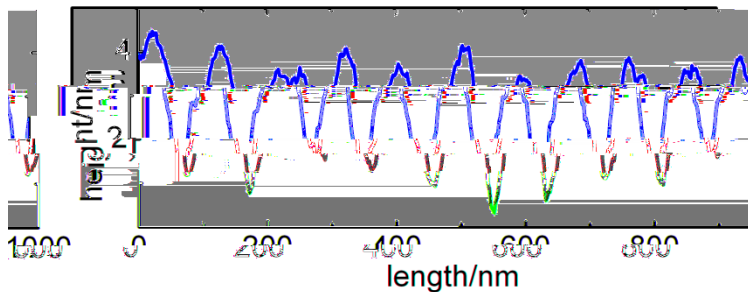
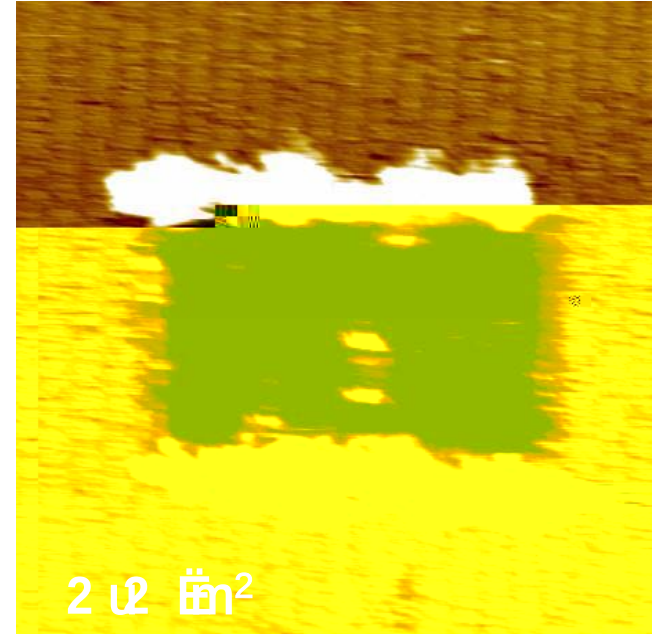
100 nm period



15 nm

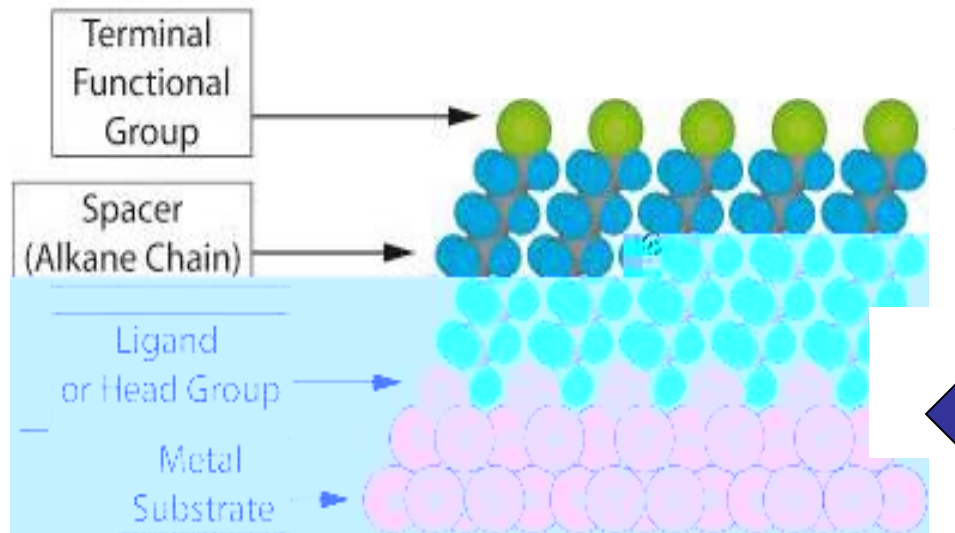
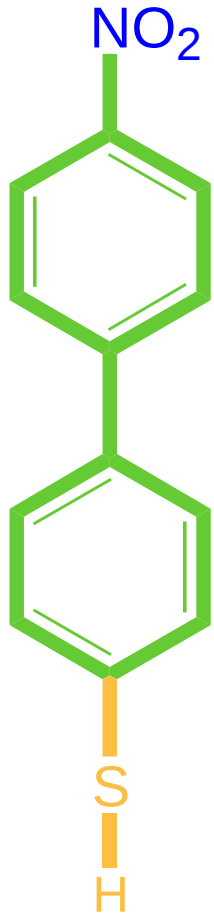


Protein lithography



Bifunctional Nanomembranes: „Janus Membranes“

SAMs have 2 functional groups!

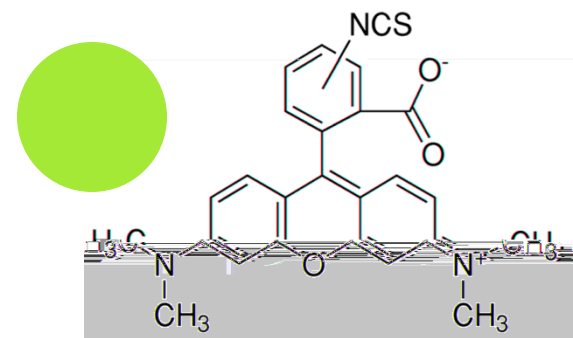
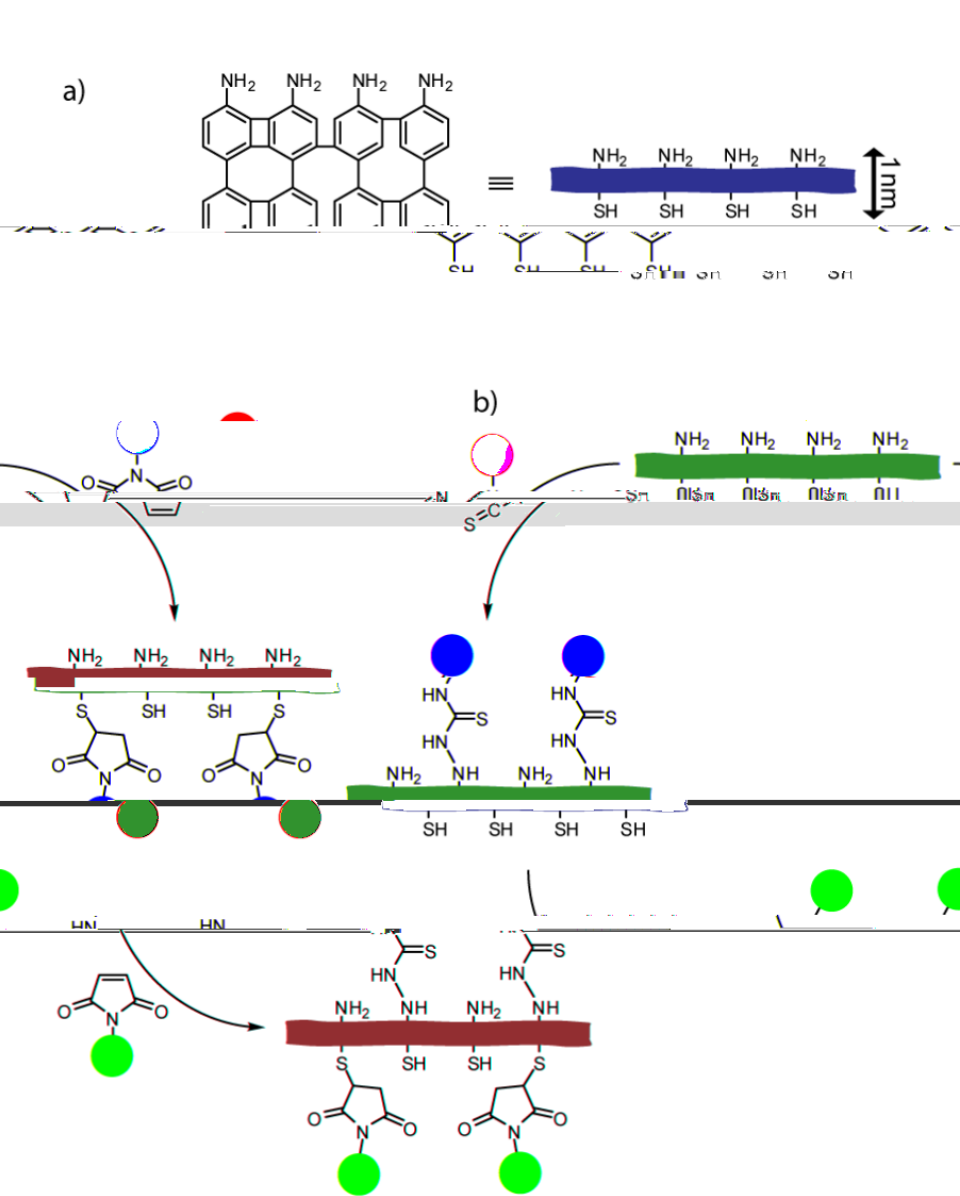


← accessible
for chemistry

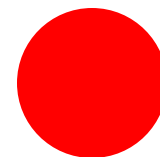
← buried

Converting the directionality of the SAM into the directionality of a 2D nanomembrane....

Different fluorescent molecules on top and bottom of membrane

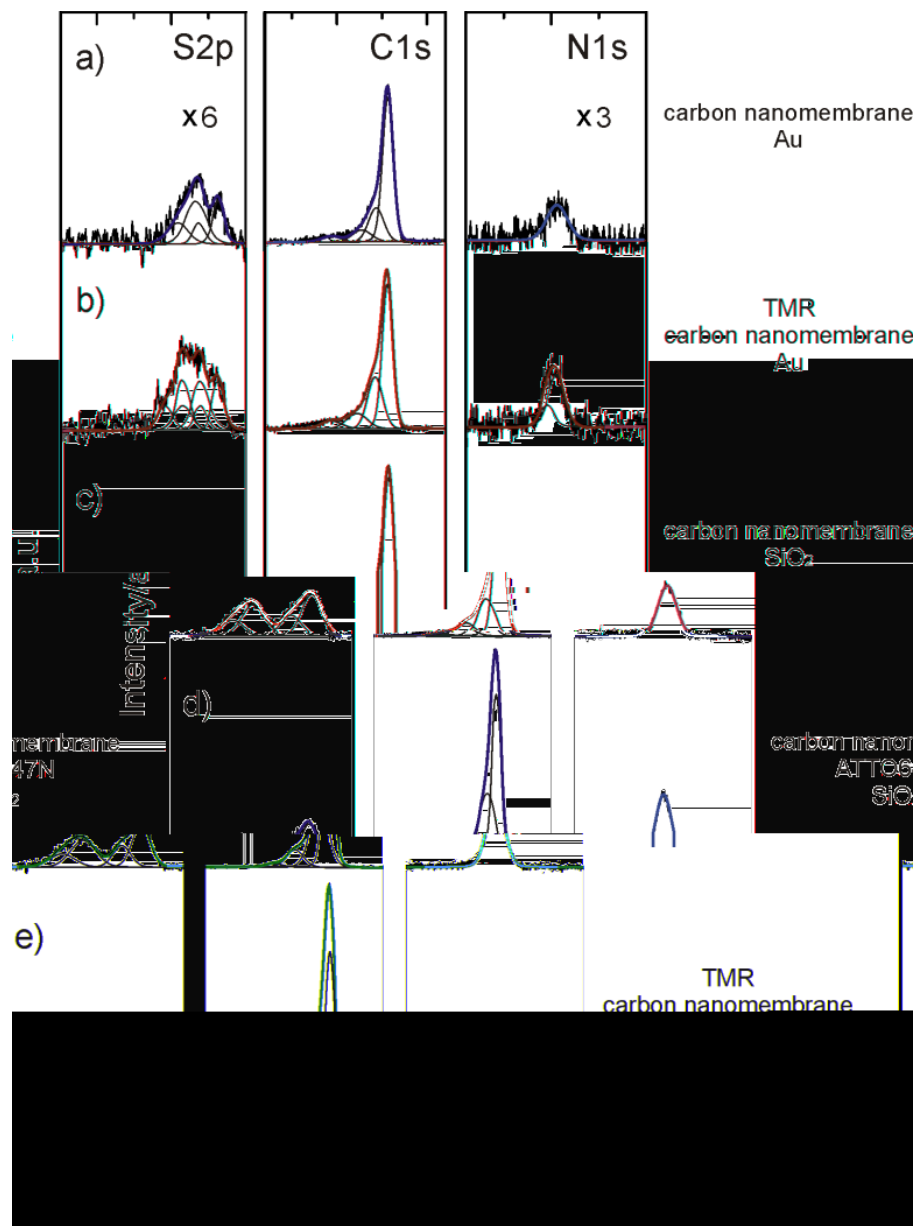
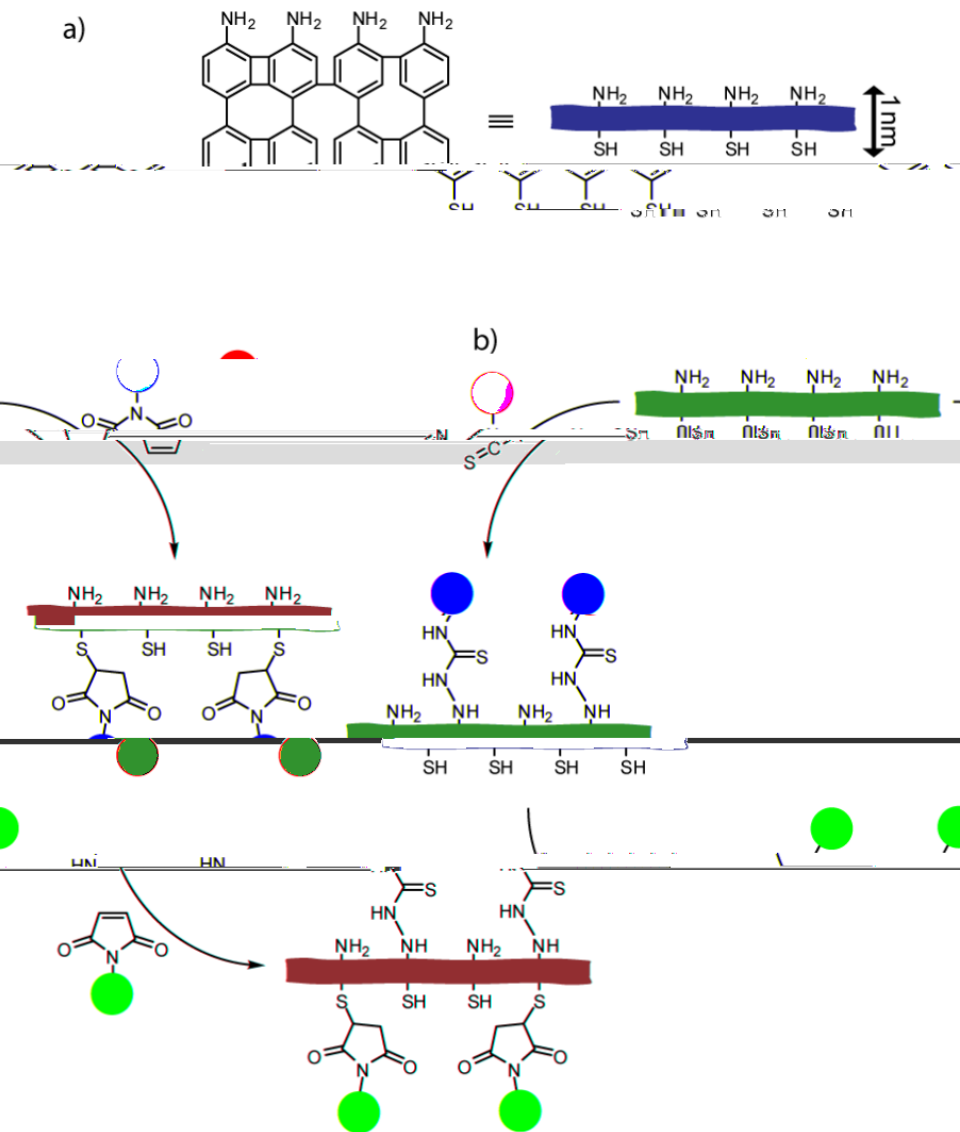


tetramethylrhodamine
isothiocyanate (TMR)

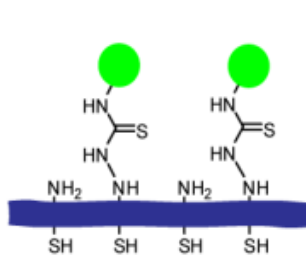


ATTO647N

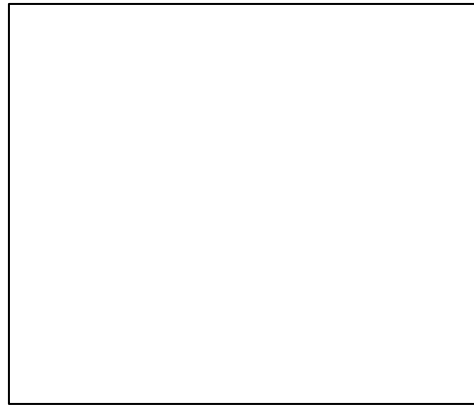
Monitoring molecular coupling by XPS



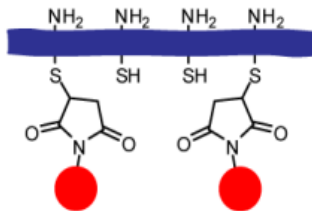
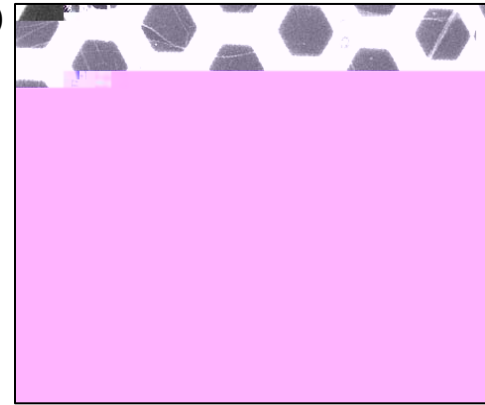
Flourescence detection of TMR and ATTO



a)



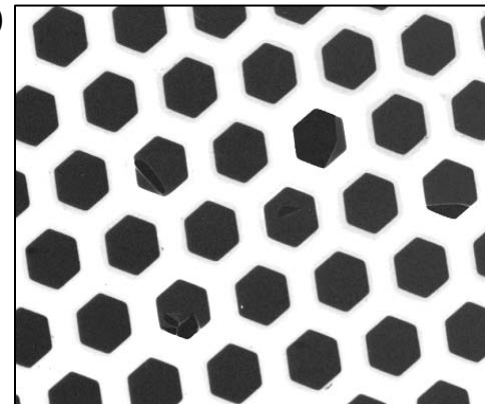
b)



c)

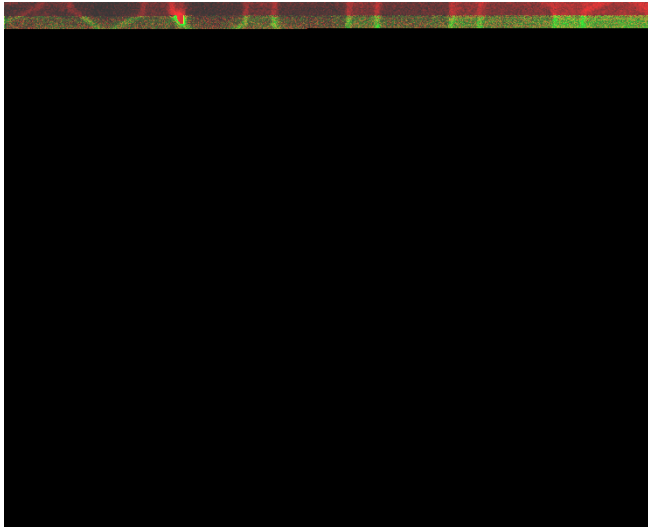


d)

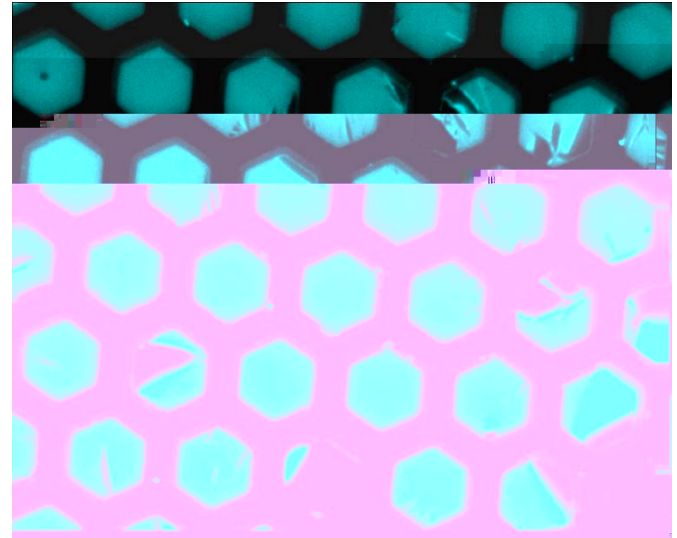


Step 3: Coupling of TMR to Top and ATTO647N to Bottom

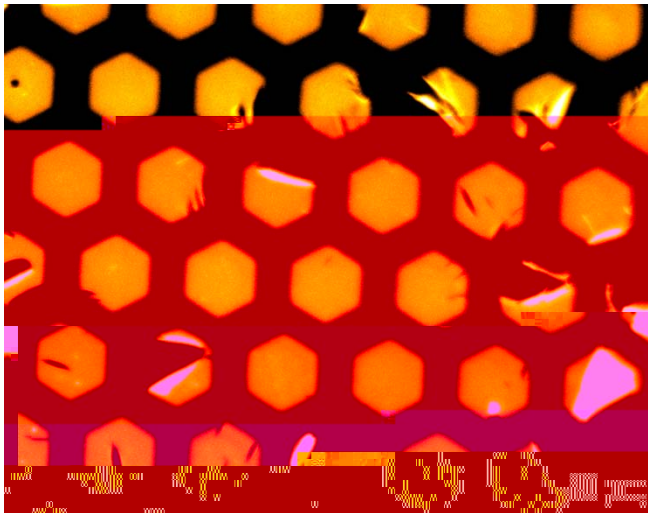
Fluorescence TMR



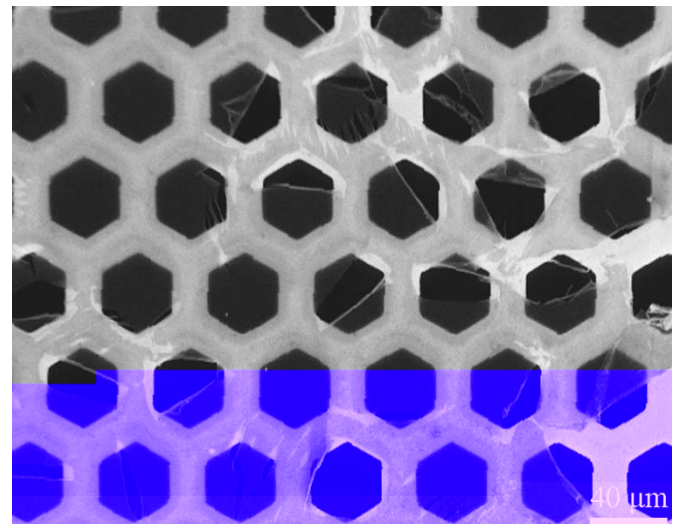
Fluorescence ATTO647N



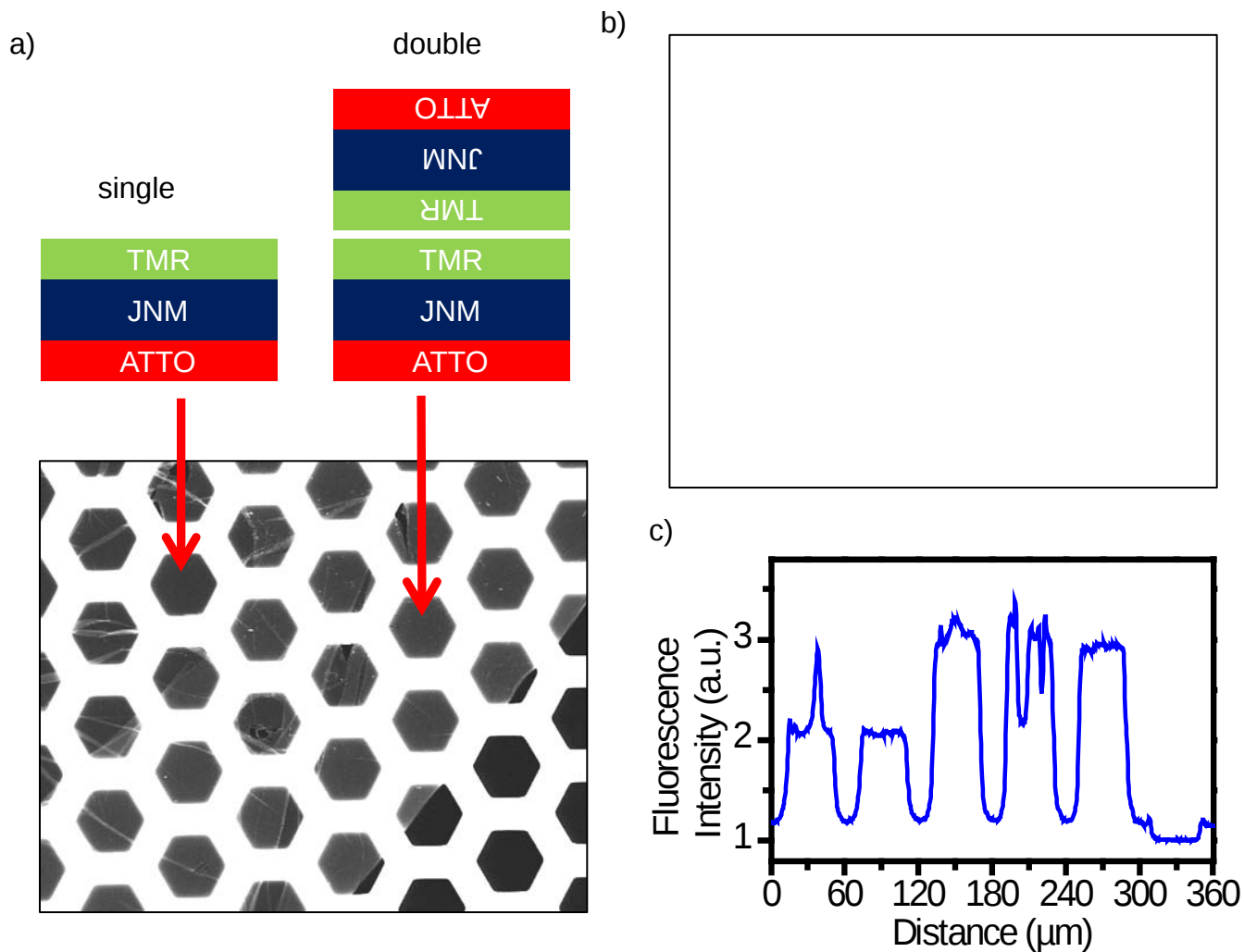
FRET (Förster Transfer)



SEM



Single and double layers





Is there a better imaging technique ?

Helium Ion Microscope

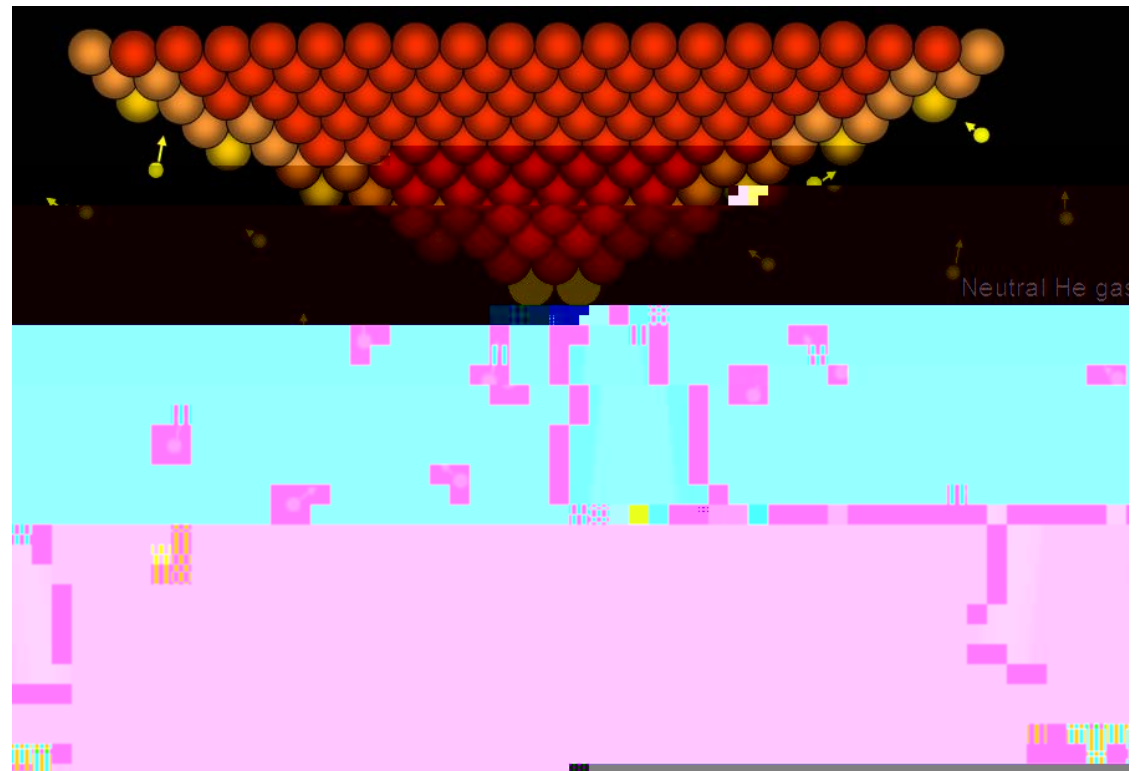


f World's first commercially available Helium Ion Microscope (Carl Zeiss)

f Analogous to a SEM but uses Helium ions instead of electrons

f Image formed using secondary electrons and backscattered ions

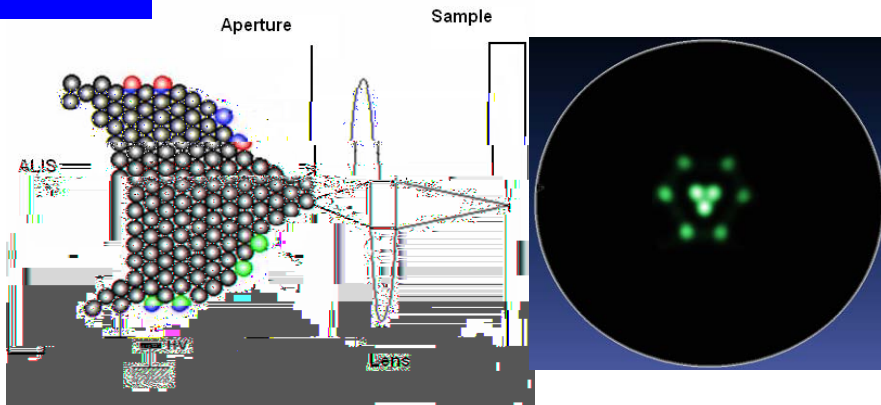
ALIS – Atomic Level Ion Source



ALIS – Atomic Level Ion Source

FIM

ALIS



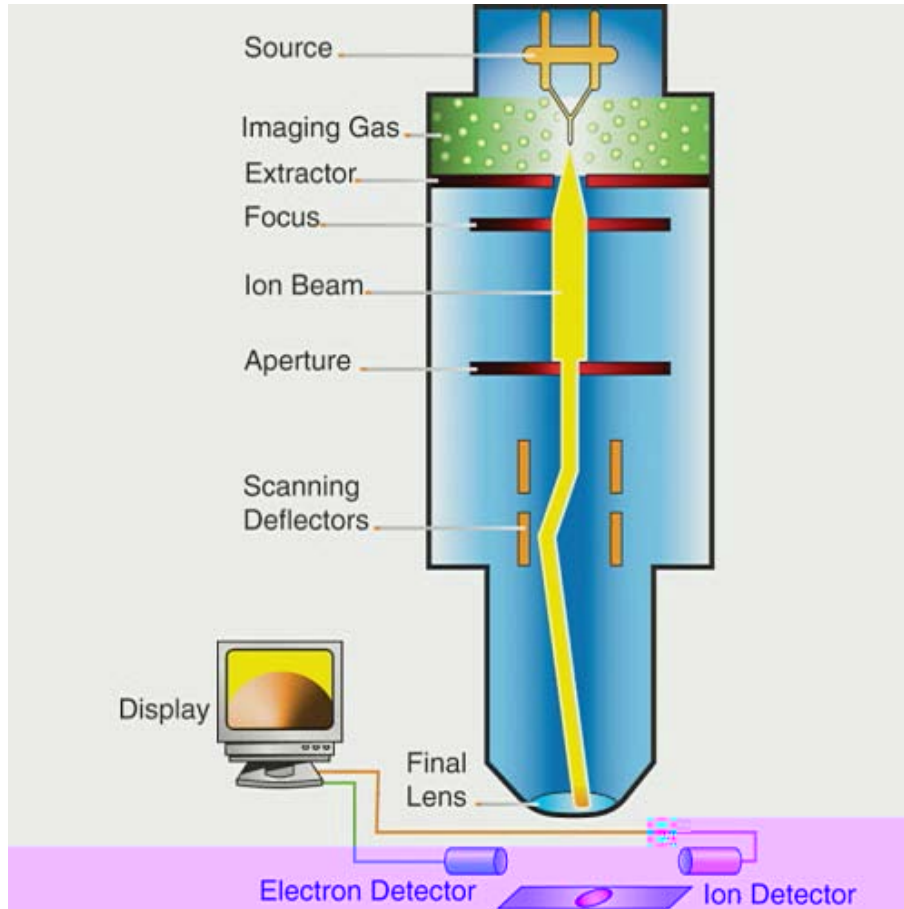
Field Ion microscope:

- f* Small emitters
- f* Beam current shared among hundreds or thousands of atom

ALIS:

- f* 3 atom shelf called the “trimer” created through field evaporation
- f* Single atom selected for final probe
- f* Source size < 1 Atom diameter

Column Architecture & Uniqueness



Architecture:

- Electrostatic optics similar to SEM / Ga FIB

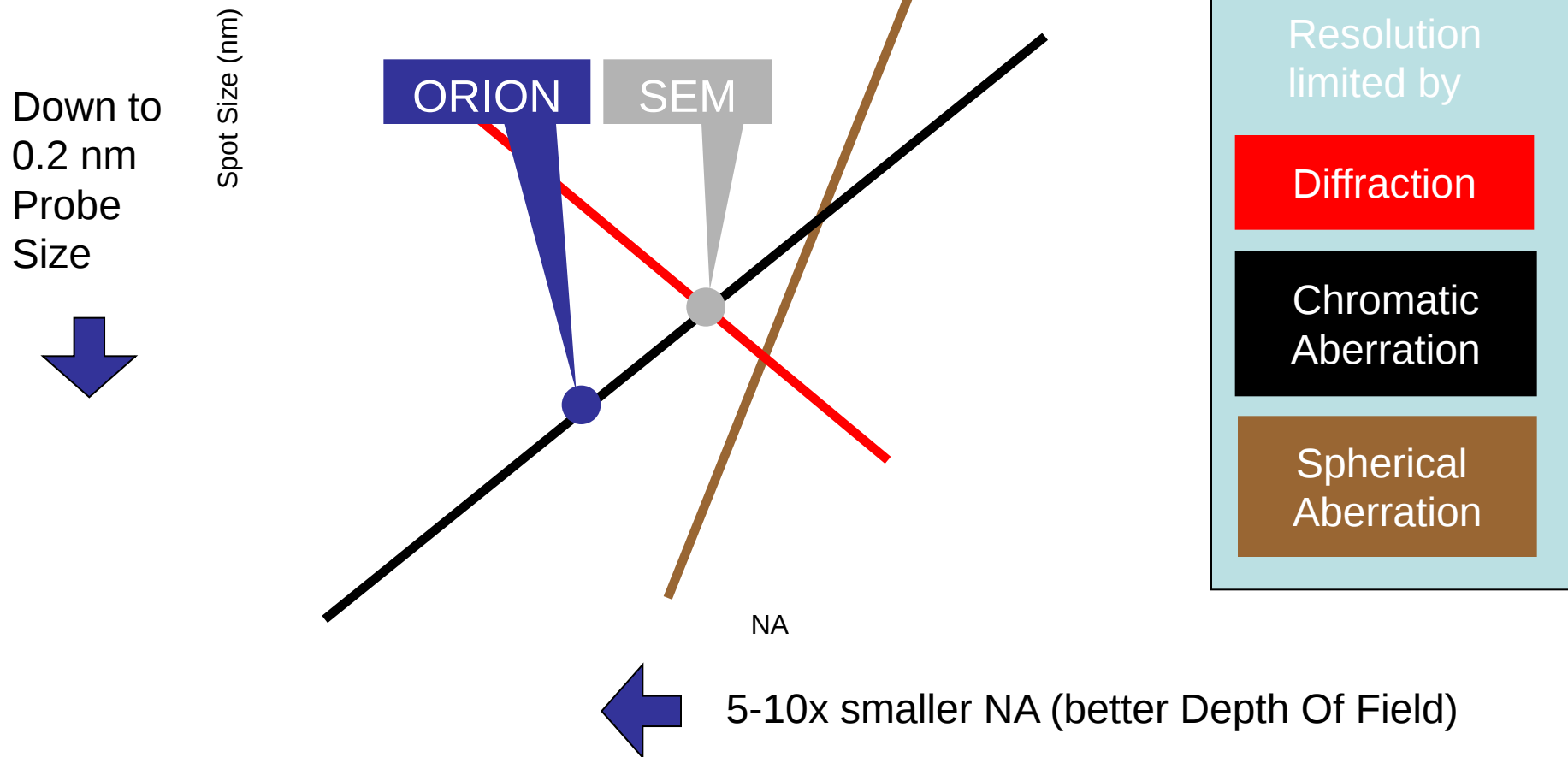
Unique:

- He Ions:
 - Wave Length (Resolution)
 - Sample Interaction
 - Contrasts
 - Surface Sensitivity
 - Charging

- Source:

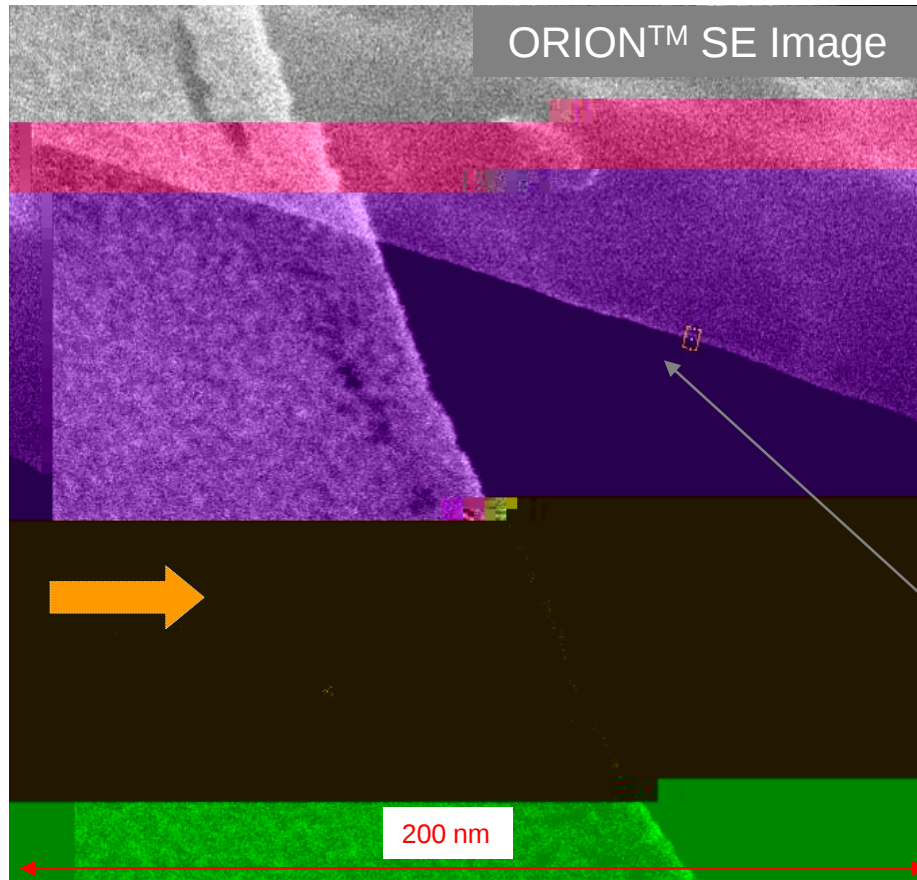
Brightness (Resolution/DoF)

ORION™ HIM Ultra High Resolution



ORION™ Resolution Recent Status Update

0.24 nm resolution demonstrated in R&D lab

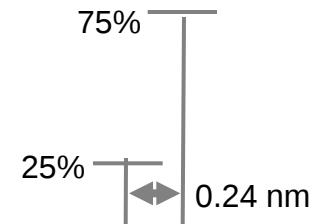


Specimen „Asbestos fibre“ on holy carbon

SE Imaging World Record
Resolution 0.24 nm (+/- 0.04 nm)

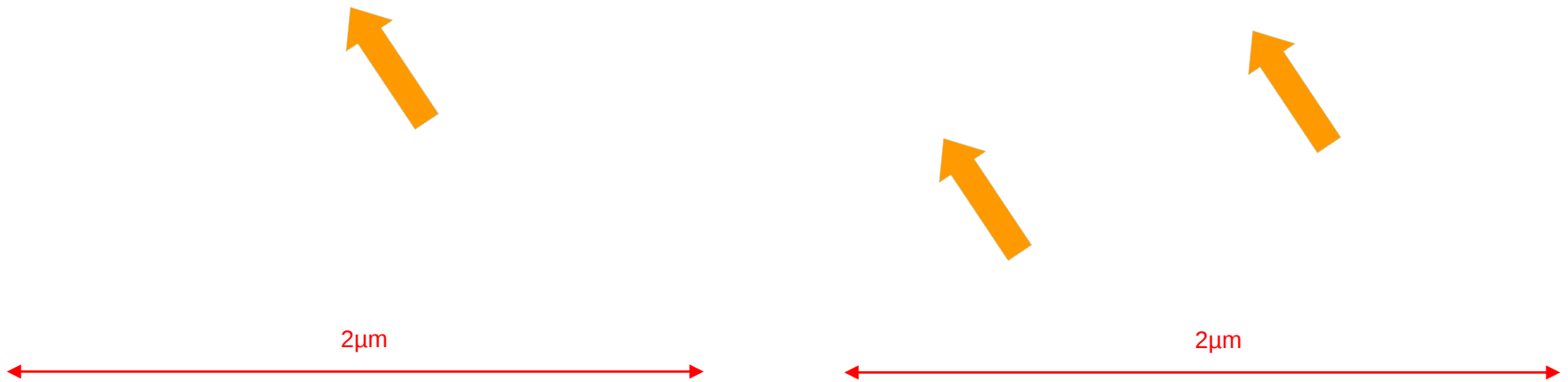
- f Working Distance: 6 mm
- f TEM like „salt and pepper pattern“ visible on carbon foil
- f 0.24 nm resolution measured repeatedly on ORION R&D System

Linescan from edge of Asbestos fibre averaged over 20 neighbouring lines



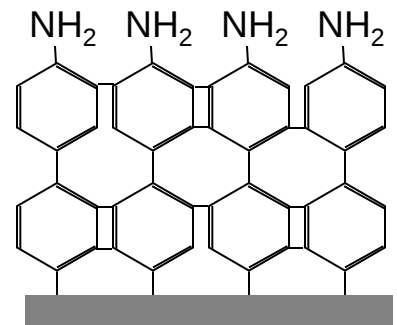
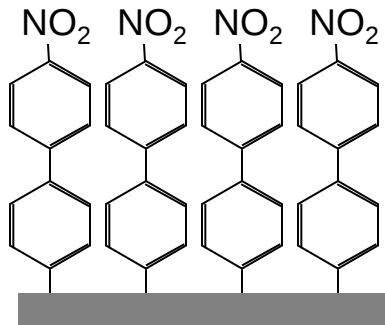
*upgrade path will be available for Orion Plus customer

Unique Material Contrast



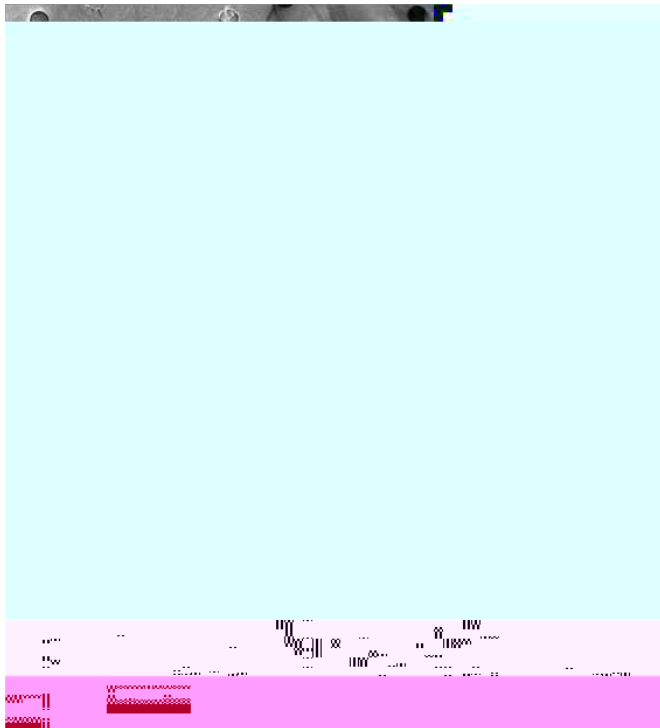
Secondary Electron image from the ORION™ shows superior material contrast in addition to surface detail

Imaging of SAM / Au Surface after Chemical Lithography

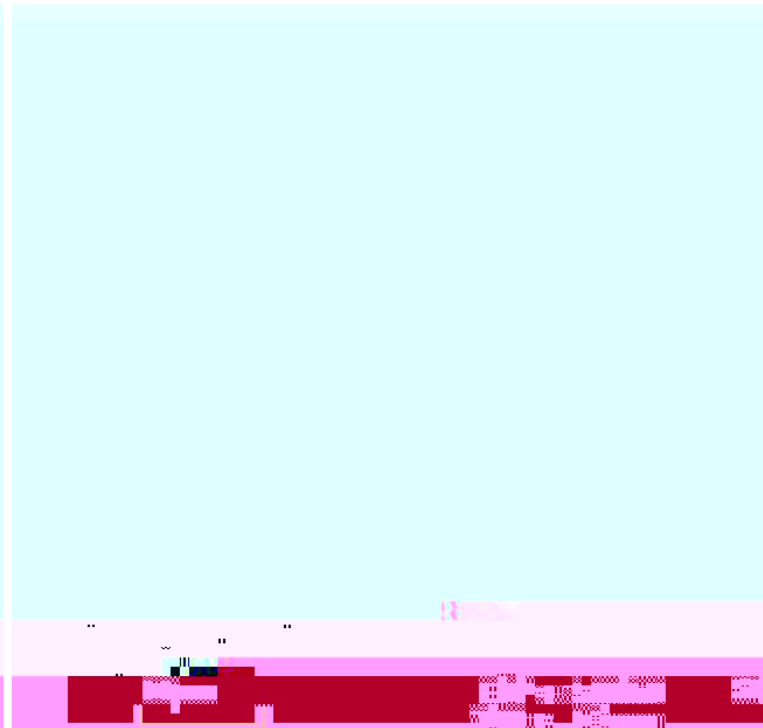


Visualizing 2-dimensional Nanomembranes

HIM



SEM



Freestanding nanomembranes on holey carbon foil

Quantifoil Holey Carbon Film, He⁺ Ion Image

(A. Beyer)

Visualizing 2-dimensional Nanomembranes

SEM

HIM



Acknowledgements:

Bielefeld group: *Andrey Turchanin, André Beyer, Christoph Nottbohm, Xianghui Zhang, Ishan Amin, Berthold Völkel, Mohamed El-Desawy, Min Ai, Eline Oberländer, Nils Mellech, Heiko Muzik, Mark Schnietz, Matthias Büenfeld, Laxman Kankate, Zinkun Zheng* <http://www.physik.uni-bielefeld.de/pss>
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Univ. Heidelberg: *Wolfgang Eck, Michael Grunze*
Univ. Frankfurt: *Helge Großmann, Robert Tampé*
KIT Karlsruhe: *Christof Wöll*
Univ. Marburg: *Gregor Witte, Daniel Rhinow, Norbert Hampp*
RWTH Aachen: *Alla Solugubenko, Joachim Mayer*
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NCEM Berkeley: *Christian Kisielowski*
Carl Zeiss: *Larry Scipioni, Frank Stietz*